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Effect of gonadotropin releasing hormone and flunixin meglumine in repeat breeder buffaloes

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Abstract

The present study was conducted to assess the efficacy of Flunixin and GnRH on conception rate among repeat breeder buffaloes which were presented to Large Animal Gynaecology Unit of Department of Veterinary Gynaecology and Obstetrics, NTR College of Veterinary Science, Gannavaram, Krishna District, Andhra Pradesh. Postpartum Graded Murrah buffaloes were screened based on history, clinico-gynaecological examination, pH of cervico-vaginal mucus, leucocyte esterase test, white side test and endometrial cytology. respectively. Based on endometrial cytology, the animals showing oestrus without subclinical endometritis (n=30) were selected and divided into three groups which were administered with normal saline (Group I, n=10), Flunixin (Group II, n=10) and Gonadotropin releasing hormone injection (Group III, n=10) between 11-13 days post AI. In the present study, higher conception rate was recorded in buffaloes injected with Gonadotropin releasing hormone – 40.00% compared to buffaloes injected with Flunixin Meglumine – 30.00%. Hence, the injection of GnRH could be recommended for field practice to improve conception rate in repeat breeder buffaloes.

Keywords: GnRH, flunixin, repeat breeder buffaloes, conception rate

1. Introduction

India is a predominantly agrarian society where lactating animals are the backbone of the national economy. Buffaloes (*Bubalus bubalis*) are premier milk-producing animals that contribute more than fifty percent of total milk production in the country (Chandra Prasad *et al.*, 2020) [8]. The term "repeat breeder" or "cyclic non-breeder" is used to characterize an animal that has been unsuccessful in conceiving after undergoing 3 or 4 services either by a fertile bull or artificial insemination (Butani *et al.*, 2016b) [5]. Despite displaying regular heat cycles and evident signs of oestrus, these animals doesn't show any clinically detectable reproductive disorders (Yusuf *et al.*, 2010) [36]. Two of the most consistent causes of repeat breeding were reduced rates of fertilization and embryonic survival (Chandra Prasad and Rao, 2014) [7]. 70-80% of embryonic mortality occurs between days 8 and 16 after insemination (Santos *et al.*, 2004) [28]. GnRH injection leads to LH secretion which causes luteinization as well as progesterone secretion. Consequently, GnRH treatments have proven successful in preventing embryonic death due to luteal deficiency (Sheldon and Dobson, 1993) [30]. An alternative and latest method to regulate maternal and fetal relation, to retard or inhibit luteolysis, is to maintain high progesterone levels in the critical days of pregnancy by application of Non-Steroid Anti-inflammatory Drugs (NSAID), leading to enhanced pregnancy rate. Non-Steroidal Anti-inflammatory Drugs (NSAID) inhibit the synthesis of cyclooxygenase (COX) enzyme resulting in the inhibition of prostaglandin production, which will protect the corpus luteum (Singh *et al.*, 2021) [31].

2. Materials and Methods

The present study was carried out on Graded Murrah buffaloes, which were presented to the Department of Animal Reproduction Gynaecology and Obstetrics, NTR College of Veterinary Science, Gannavaram from November 2022 to October 2023. Postpartum Graded Murrah buffaloes with different parities were screened (n=71) based on the following criteria *viz.*, history, clinico-gynaecological examination, pH of cervico-vaginal mucus, LEST, white side test and uterine discharge cytology.

Blood samples were collected in EDTA vacutainers to assess the haematological parameters, while blood collected in clot-activating vacutainers was utilized to obtain the necessary serum for assessing biochemical parameters. Haemoglobin levels were determined and expressed in grams per deciliter (g/dL) using the Sahli's haemoglobinometer technique, while packed cell volume (PCV) was assessed using a microhematocrit scale and reported as a percentage. Biochemical parameters, such as Total protein (Biuret and end point assay method) and Cholesterol (CHOD-PAP enzymatic end point assay method) and hormones such as Cortisol and Progesterone evaluated by Calbiotech ELISA kit. Selected repeat breeder buffaloes which were devoid of palpable reproductive abnormalities and negative for subclinical endometritis were divided into three groups with each Group 10. Group I were administered with normal saline, Group II were injected with 1.1 mg/Kg b.wt. of flunixin meglumine and Group III were injected with 10 µg of GnRH (Receptal) intramuscularly, respectively. All groups were administered once between days 11-13 post AI. The statistical analysis of the data was done as per the procedures outlined by Snedecor and Cochran (1994) [32]. The haematological and serum biochemical parameters within and between the groups were compared by Independent Sample T Test and One Way ANOVA. The conception rate was analysed by Fisher exact test.

3. Results and Discussion

3.1 Haematological parameters

In the present study the mean haemoglobin (gm/dL) and PCV (%) concentration recorded was non-significant ($p>0.05$) in between conceived and non-conceived repeat breeding buffaloes. The mean haemoglobin (gm/dL) concentration recorded was non-significantly ($p>0.05$) higher in conceived compared to non-conceived repeat breeding buffaloes of Group I (8.59 ± 0.03 vs 8.52 ± 0.04), Group II (8.63 ± 0.10 vs 8.51 ± 0.04) and Group III (8.64 ± 0.05 vs 8.54 ± 0.06), respectively. The mean PCV (%) level recorded was non-significantly ($p>0.05$) higher in conceived compared to non-conceived repeat breeding buffaloes of Group I (32.50 ± 0.65 vs 31.88 ± 0.80), Group II (34.30 ± 0.41 vs 32.02 ± 0.60) and Group III (34.50 ± 0.62 vs 33.01 ± 0.93), respectively, which were similar to reports of Patel *et al.* (2016) [24], who also found a non-significantly higher values of Hb (gm/dL) and PCV (%) in conceived compared to non-conceived buffaloes (12.36 ± 0.85 vs 11.41 ± 0.82) (34.05 ± 1.39 vs 32.41 ± 1.63) (Table 1, Plate 1). The higher concentrations of HB and PCV were desirable physiological characteristics for efficient transport of oxygen and carbon-di-oxide which is essential for maintaining the health of animals. Consequently, buffaloes exhibiting higher concentrations of HB and PCV were more economical due to enhanced reproductive efficiency.

3.2 Biochemical parameters

In the present study, no significant ($p>0.05$) difference was recorded in the mean concentration of total protein (gm/dL) between conceived and non-conceived buffaloes. The mean serum total protein (gm/dL) in conceived buffaloes of Group I, II and III was 7.28 ± 0.08 , 7.57 ± 0.20 and 7.68 ± 0.08 , meanwhile the same in non-conceived buffaloes was 7.16 ± 0.09 , 7.33 ± 0.08 and 7.43 ± 0.05 , respectively, which were in accordance to Savalia *et al.* (2014) [29] and Chirag *et al.* (2016) [9]. The results might be attributed to better feeding of animals which increased the concentration of amino acids required for biosynthesis of gonadotropins and other gonadal

hormones (Table 2, Plate 2).

The serum total cholesterol levels were non-significant ($p>0.05$) between conceived and non-conceived buffaloes of the Group I and II with higher values in conceived buffaloes. The mean cholesterol concentration (mg/dL) was recorded as 84.72 ± 0.07 , 82.93 ± 0.66 and 82.71 ± 0.62 in conceived buffaloes of the Group I, II and III, while the same in non-conceived buffaloes was 83.70 ± 0.89 , 82.24 ± 0.65 and 81.70 ± 0.57 , respectively. (Table 2, Plate 2). The present observations were also very much similar to that documented by Chirag *et al.* (2016) [9] who observed insignificantly higher mean cholesterol concentration in conceived than non-conceived repeat breeder buffaloes (142.32 ± 18.84 vs. 140.73 ± 26.96). In the present study, the non-significantly higher levels of cholesterol in conceived compared to non-conceived repeat breeding buffaloes across all groups might be due to extraction of cholesterol stored in the tissues and variations in quality of feed, energy status of the animal and the level of milk production (Ashwani *et al.* 2015) [3].

3.3 Hormonal Profiles

The mean serum cortisol levels in conceived buffaloes of Group I, II and III was 6.02 ± 0.06 , 6.04 ± 0.18 and 6.02 ± 0.11 ng/mL whereas in non-conceived buffaloes, the levels were 6.15 ± 0.04 , 6.14 ± 0.09 and 6.12 ± 0.10 , respectively (Table 2, Plate 2). Rajamanickam *et al.* (2022) [25] observed a significant increase in cortisol levels in non-pregnant animals than pregnant animals suggesting that systemic cortisol levels may affect embryo implantation and hormonal balance. The variations in the serum cortisol levels of the present study might be due to environmental stress (Agarwal and Sharma 2002) [2], like hot temperature and less stable ventilation management which interferes with the release of hormones there by effecting fertility (Da Costa *et al.*, 2017) [10].

Among the Group I buffaloes, the mean serum progesterone concentration (ng/mL) on 0, 11th and 42nd day of insemination was recorded as 0.13 ± 0.01 , 3.29 ± 0.15 and 4.68 ± 0.16 respectively in conceived buffaloes, whereas in non-conceived buffaloes it was 0.12 ± 0.02 , 2.31 ± 0.08 and 1.39 ± 0.07 . The mean serum progesterone levels in Group I conceived buffaloes were in close agreement with Pandey *et al.* (2016) [23] as 4.76 ± 0.38 , whereas lower value was recorded by Vijayarajan *et al.* (2007) [34] as 3.85 ± 0.05 and higher value by Mandal *et al.* (2009) [20] as 5.34 ± 0.82 (ng/mL). The mean serum progesterone (ng/mL) in Group II on the day of estrus was 0.17 ± 0.43 and 0.14 ± 0.01 in conceived and non-conceived buffaloes, while the same on 11th day was 3.63 ± 0.14 and 2.64 ± 0.08 , whereas on 42nd day of post breeding was 5.49 ± 0.10 and 1.64 ± 0.07 , correspondingly. The mean serum progesterone levels in conceived buffaloes of Group II was in consonance with Rokade (2015) [26] and Damarany and Ghanem (2020) [11] who recorded the values as 6.61 and 6.72, correspondingly, whereas on contrary, lower values were recorded by Neto *et al.* (2008) [22] as 0.44 ng/mL. On the other hand, Rossetti *et al.* (2011) [27] and Barkhori *et al.* (2018) [4] reported higher value as 9.78 and 9.21 ng/mL, respectively. The mean serum progesterone concentration was recorded in conceived buffaloes of Group III on the day of oestrus, 11th and 42nd day of post breeding was 0.31 ± 0.06 , 4.12 ± 0.17 and 6.82 ± 0.31 ng/mL, while the corresponding levels in non-conceived buffaloes was 0.19 ± 0.02 , 3.45 ± 0.10 and 1.93 ± 0.17 ng/mL, respectively (Table 3, Plate 3). The mean serum progesterone levels on 0th day shows non-significant difference ($p>0.05$) between conceived and non-conceived buffaloes between three groups, whereas on 11th

and 42nd day the serum progesterone level is significantly higher ($p < 0.05$) in conceived buffaloes of Group III than Group II and Group I. Campanile *et al.* (2008) [6], Kishorekumar (2010) [18] and Deshpande (2017) [13] observed increased progesterone levels in conceived buffaloes compared to non-conceived buffaloes following GnRH injection. The values in Group III conceived buffaloes were in agreement with Mandal *et al.* (2009) [20], Lattoo *et al.* (2013) [19] and Hemadepthi (2020) [17] as 7.33 ± 1.10 , 6.66 , and 6.21 ± 0.09 (ng/mL). The increased progesterone levels in Group III might be due the effect of GnRH injection which might have increased the life span of CL due to its luteotropic and luteoprotective properties thereby contributing to pregnancy maintainance (Lattoo *et al.*, 2013) [19].

3.4 Conception Rate

The conception rate in Group I buffaloes was 10.00% which is in close agreement with Kishorekumar (2010) [18] as 15.00 per cent. The conception rate was 30.00% in Group II buffaloes of the present study was in close agreement to the reports Damarany *et al.* (2017) [12] and Barkhori *et al.* (2018) [4] as 33.00 and 35.00 per cent, correspondingly. On contrary, higher conception rates was recorded by Geary *et al.* (2010) [14], Rossetti *et al.* (2011) [27], Navrange *et al.* (2012) [21] and

Rokade (2015) [26] as 58.00, 63.00, 66.00 and 75.00 per cent respectively, which is due to different days and routes of administration. The present study, recorded a higher conception rate 40.00% (Table 4, Plate 4) in Group III buffaloes on 11th day of post breeding which was in close agreement to the findings of Vijayarajan *et al.* (2007), [34] Gumen *et al.* (2011) [15] and Venkateswarlu (2019) [33] who observed the conception rates as 40.00, 44.30 and 46.67 per cent. On the contrary, Abo-Farw *et al.* (2016) [1] and Zakiuddin *et al.* (2022b) [37] recorded the conception rates as 80.00 and 90.00 per cent respectively in buffaloes administered with GnRH analogues. Research studies have demonstrated that administering GnRH on day 5 or 11 (Willard *et al.*, 2003) [35] and between days 11 to 14 (Hansen 2002) [16], following artificial insemination results in increased serum concentration of progesterone and a tendency towards higher pregnancy rates. Embryonic mortality is one of the cause of repeat breeding which may be due to luteal insufficiency in this group it was cared by supplementing the exogenous GnRH on days 11-13 of oestrus cycle, which might had reduced oestradiol-17 β secretion during maternal recognition of pregnancy (Lattoo *et al.*, 2013) [19] and it may be the possible reason for higher conception rate in Group II of the present study.

Table 1: Haematological parameters in conceived and non-conceived buffaloes in different groups

Haematological parameters		Groups		
		Group I	Group II	Group III
Haemoglobin (g/dL)	Conceived	8.59 \pm 0.03	8.63 \pm 0.10	8.64 \pm 0.05
	Non-conceived	8.52 \pm 0.04	8.51 \pm 0.04	8.54 \pm 0.06
PCV (%)	Conceived	32.50 \pm 0.65	34.30 \pm 0.41	34.50 \pm 0.62
	Non-Conceived	31.88 \pm 0.80	32.02 \pm 0.60	33.01 \pm 0.93

Table 2: Total protein (g/dL), Cholesterol (mg/dL) and Cortisol (ng/mL) levels between conceived and non-conceived buffaloes of different groups.

Groups	Conceived /Non-conceived	Total protein (g/dL)	Cholesterol (mg/dL)	Cortisol (ng/mL)
Group I	Conceived	7.28 \pm 0.08	84.72 \pm 0.07	6.02 \pm 0.06
	Non-conceived	7.16 \pm 0.09	83.70 \pm 0.89	6.15 \pm 0.04
Group II	Conceived	7.57 \pm 0.20	82.93 \pm 0.66	6.04 \pm 0.18
	Non-Conceived	7.33 \pm 0.08	82.24 \pm 0.65	6.14 \pm 0.09
Group III	Conceived	7.68 \pm 0.08	82.71 \pm 0.62	6.02 \pm 0.11
	Non-Conceived	7.43 \pm 0.05	81.70 \pm 0.57	6.12 \pm 0.10

Table 3: Serum Progesterone (ng/mL) in conceived and non-conceived buffaloes in different groups

Group	Status	Days after AI		
		0	11	40-42
Group I	Conceived	0.13 \pm 0.01	3.29 \pm 0.15*	4.68 \pm 0.16*
	Non Conceived	0.12 \pm 0.02	2.31 \pm 0.08*	1.39 \pm 0.07*
Group II	Conceived	0.17 \pm 0.43	3.63 \pm 0.14*	5.49 \pm 0.10*
	Non Conceived	0.14 \pm 0.01	2.64 \pm 0.08*	1.64 \pm 0.07*
Group III	Conceived	0.31 \pm 0.06	4.12 \pm 0.17*	6.82 \pm 0.31*
	Non Conceived	0.19 \pm 0.02	3.45 \pm 0.10*	1.93 \pm 0.17*

* Indicates significance difference ($p < 0.05$) in level of progesterone (ng/mL) between rows and columns of conceived and non-conceived buffaloes of different groups on 11th and 40-42nd day.

Table 4: Comparison of conception rate by Fisher exact test

Groups		No. of animals conceived	Fisher exact test statistic value
Group I	Control	1	0.58
Group II	Flunixin	3	
Group I	Control	1	
Group III	GnRH	4	0.30
Group II	Flunixin	3	
Group III	GnRH	4	
			1.0

Non-significant difference ($p > 0.05$) was observed between the Groups I, II and III for conception rates.

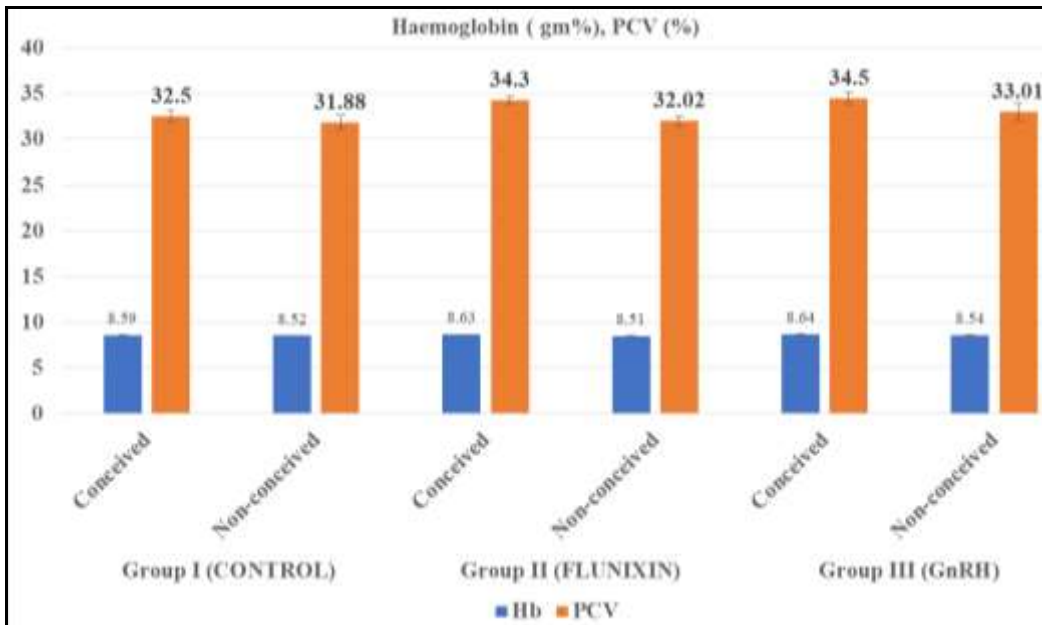


Plate 1: Haemoglobin (g/dL) and PCV (%) in conceived and non-conceived buffaloes of different groups

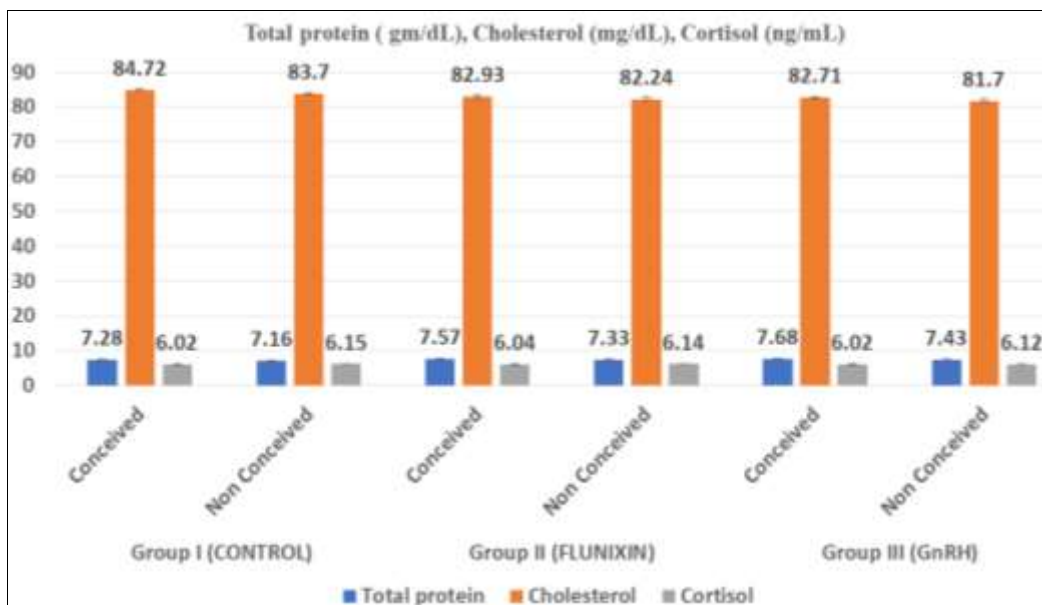


Plate 2: Total protein (gm/dL), Cholesterol (mg/dL), Cortisol (ng/mL) in conceived and non-conceived buffaloes of different groups

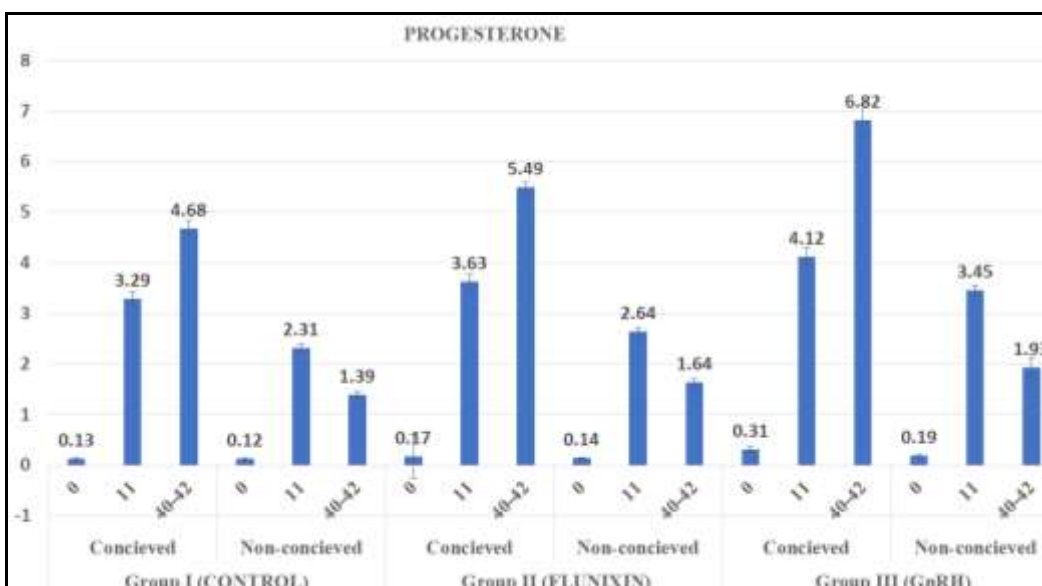


Plate 3: Levels of progesterone (ng/mL) in conceived and non-conceived buffaloes of different groups

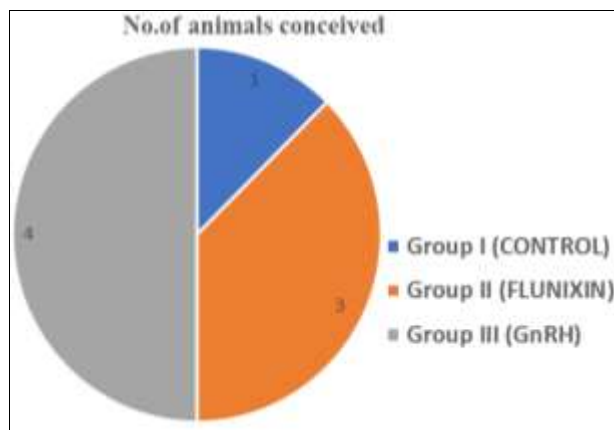


Plate 4: Conception rate in percentage in Group I, II and III

4. Conclusion

The current study concludes that GnRH injection could be a viable recommendation for practical application in the field to enhance conception rates in non-infectious repeat breeder buffaloes.

5. References

1. Abo-Farw MA, El-Ratel IT, Abdel-Khalek AE, Abouelghet HA, Ghoneim MM. Effect of GnRH treatment during different times post-mating on pregnancy rate of buffalo heifers. *Egyptian Journal of Animal Production*. 2016;53:59-64.
2. Agarwal RG, Sharma IJ. Levels of serum free thyroid hormones and cortisol in estrus, anoestrus and repeat breeding buffaloes. *The Indian Journal of Animal Reproduction*. 2002;23:73-74.
3. Ashwani K, Sharma U, Singh R, Kumar S, Kumar S. Changes in hemato-biochemical profile in postpartum anoestrus Murrah buffaloes subjected to different hormonal protocols. *Indian Veterinary Journal*. 2015;92:50-52.
4. Barkhori -Mehni S, Karami-Shabankareh H, Masoumi R, Kazemi-Bonchenari M, Pezeshki A, Badiei A, *et al.* Effect of exogenous progesterone or flunixin meglumine after AI on serum progesterone concentration and pregnancy per AI in lactating dairy cows. *Animal Reproduction*. 2018;15:140.
5. Butani M, Dhama AJ, Shah RG, Sarvaiya NP, Killedar A. Management of repeat breeding in buffaloes under field conditions using hormonal and antibacterial therapies. *Buffalo Bulletin*. 2016b;35:83-92.
6. Campanile G, Vecchio D, Neglia G, Di Palo R, Prandi A, D'occhio MJ. Progesterone and pregnancy status of buffaloes treated with a GnRH agonist. *Livestock science*. 2008;115:242-248.
7. Prasad CB, Rao KA. Effect of administration of busserelin acetate on different days of oestrous cycle in graded murrah buffaloes. *International Journal of Recent Scientific Research*. 2014;5:1558-1559.
8. Chandra Prasad B, Naidu V, Srinivas M, Raganath M, Kumar A. Evaluation of different hormonal protocols in postpartum anestrus buffaloes under farm and field during breeding and low breeding seasons. *Journal of Pharma Innovation*. 2020;9:210-218.
9. Chirag PP, Patel DM, Dhama A, Hadiya KK, Patel J, Buhecha KV. Effect of ovsynch and mid-cycle PGF 2α treatment protocols on conception rates and plasma biochemical and minerals profile in repeat breeding cows and buffaloes. *International Journal of Advanced Veterinary Science and Technology*. 2016;5:217-225.
10. Da Costa Freitas J. Cortisol Hormones Profiles of Repeat Breeding Local Cattle. In *International Seminar on Tropical Animal Production*; c2017. p. 799-803.
11. Damarany AI, Ghanem N. Effect of flunixin meglumine and aspirin administration on conception rate and estrous cycle characteristics of Egyptian Baladi cows during hot season. *Tropical Animal Health and Production*. 2020;52:2969-2976.
12. Damarany DI. Influence of season of calving, parity and flunixin meglumine administration on conception rate in repeat breeder Egyptian Baladi Cows and their crosses. *Journal of Animal and Poultry Production*. 2017;8:409-414.
13. Deshpande VP. Augmentation of fertility with luteotrophic hormones in repeat breeding buffaloes. M.V.Sc thesis submitted to Maharashtra Animal Fishery Sciences University, Nagpur; c2017.
14. Geary TW, Ansotegui RP, MacNeil MD, Roberts AJ, Waterman RC. Effects of flunixin meglumine on pregnancy establishment in beef cattle. *Journal of Animal Science*. 2010;88:943-949.
15. Gumen, Keskin A, Yilmazbas-Mecitoglu G, Karakaya E, Cevik S, Balci F. Effects of GnRH, PGF 2α and oxytocin treatments on conception rate at the time of artificial insemination in lactating dairy cows. *Czech Journal of Animal Science*. 2011;56:279-283.
16. Hansen PJ. Embryonic mortality in cattle from the embryos perspective *Journal of Animal Science*. 2002;80:33-44.
17. Hemadeepthi M. Therapeutic Efficacy of PGF 2α AND GnRH in repeat breeder graded buffaloes. MVSc thesis submitted to Sri Venkateswara Veterinary University, Tirupati; c2020.
18. Kishorekumar S. Modulation of conception rate and progesterone concentration in repeat breeding graded murrah buffaloes by using GnRH analogue and hCG. M.V.Sc thesis submitted to Sri Venkateswara Veterinary University, Hyderabad; c2010.
19. Lattoo MZ, Prasad S, Gupta HP, Hussain, A. Effect of Post-Mating GnRH Treatment on Serum Progesterone Profile and Conception Rate in Buffaloes. *Iranian Journal Of Applied Animal Sciences*. 2013;3:83-90.
20. Mandal DD, Srivastava SK, Kumar P. Effect of GnRH during various stages of estrus cycle on fertility and plasma progesterone in buffaloes. *The Indian Journal of Animal Reproduction*. 2009;30:23-27.
21. Navrange PP, Ingawale MV, Pawshe CH, Deshmukh SG, Munde VK. Efficacy of COX-2 inhibitor on conception rate in buffaloes. *The Indian Journal of Animal Reproduction*. 2013;33:64-66.
22. Neto AP, deLucca FM, Alberton J, FalciMota M, Lucacin E, Borges AM, *et al.* Assessment on the effects of flunixin meglumine on the serum concentration of progesterone and the estrous cycle in crossbred heifers and cows. 2008;15:10-14.
23. Pandey NKJ, Gupta HP, Prasad S, Sheetal SK. Plasma progesterone profile and conception rate following exogenous supplementation of gonadotropin-releasing hormone, human chorionic gonadotropin, and progesterone releasing intra-vaginal device in repeat-breeder crossbred cows. *Veterinary world*. 9:559.
24. Patel MD, Lateef A, Das H, Patel AS, Patel AG, Joshi AB. Effect of age, sex and physiological stages on hematological indices of Banni buffalo (*Bubalus*

- bubalis*). *Veterinary World*. 2016;9:38.
25. Rajamanickam K, Ali MS, Leela V. Effect of systemic cortisol on pregnancy rate in repeat breeding cows during early pregnancy. *Indian Journal of Animal Research*. 2022;56:1454-1461.
 26. Rokade PM. Efficacy of cyclooxygenase (cox) inhibitors in noninfectious repeat breeder cows. M. V. Sc. thesis submitted to Maharashtra Animal and Fisheries Sciences University, Nagpur; c2015.
 27. Rossetti RC, Perdigão A, Mesquita FS, Sá Filho M, Nogueira GDP, Machado R, *et al.* Effects of flunixin meglumine, recombinant bovine somatotropin and/or human chorionic gonadotropin on pregnancy rates in Nelore cows. *Theriogenology*. 2011;76:751-758.
 28. Santos JE, Thatcher WW, Chebel RC, Cerri RL, Galvao KN. The effect of embryonic death rates in cattle on the efficacy of estrous synchronization programmes. *Animal Reproduction Sciences*. 2004;82:513-535.
 29. Savalia KK, Dhama AJ, Hadiya KK, Patel KR, Sarvaiya NP. Influence of controlled breeding techniques on fertility and plasma progesterone, protein and cholesterol profile in true anestrus and repeat breeding buffaloes. *Veterinary World*. 2014;7:727-732.
 30. Sheldon IM, Dobson H. Effects of gonadotrophin releasing hormone administered 11 days after insemination on the pregnancy rates of cattle to the first and later services. *The Veterinary Record*. 1993;133:160-163.
 31. Singh SP, Ankesh Kumar A, Bhavsar P, Bhavsar M, Sourya N, Singh AK, *et al.* Application of non-steroidal anti-inflammatory drugs (NSAIDs) for improvement of cattle fertility. *Pharma Innovation*. 2021;10:404-407.
 32. Snedecor GM, Cochran WC. *Statistical Methods*. 9th edition Oxford and IBM Publishing Company. Mumbai, India; c1994. p. 124-165.
 33. Venkateswarlu M. Studies on efficacy of GnRH and hCG during mid-luteal stage of cycle in repeat breeder graded Murrah buffaloes. MVSc thesis submitted to Sri Venkateswara Veterinary University, Tirupati; c2019.
 34. Vijayarajan A, Chandrahasan C, Ezakial Napoleon RZ. Effect of pre and post insemination substitution of GnRH in repeat breeding buffaloes. *Indian Veterinary Journal*. 2007;84:940-943.
 35. Willard S, Gandy S, Bowers S, Graves K, Elias A, Whisnant C. The effects of GnRH administration post insemination on serum concentrations of progesterone and pregnancy rates in dairy cattle exposed to mild summer heat stress. *Theriogenology*. 2003;59:1799-1810.
 36. Yusuf M, Nakao T, Ranasinghe RB, Gautam G, Long ST, Yoshida C, *et al.* Reproductive performance of repeat breeders in dairy herds. *Theriogenology*. 2010;73:1220-1229.
 37. Zakiuddin M, Tandle MK, Usturge SM, Patil NA, Kumar DD, Kasaralikal VR, *et al.* Therapeutic management of non-infectious repeat breeder buffaloes using GNRH analogue; c2002b. p. 2061-2065.