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Assessment and utilization of tree and shrub forages as livestock feed in Kenya: A review

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Abstract

The nutritive value of indigenous multipurpose trees and shrubs is reviewed. The ten local Kenyan MPTS contain appreciable crude protein (112-321 gkg⁻¹ DM) total extractable phenolics (4.5- 52.3 gkg⁻¹ DM), total extractable tannins (0.6–38.5 g/kg DM) and major and microelements. Although tannins may impair nutrient utilization, they contribute to bypass protein and ameliorate rumen methane production. The ranking of these tree forages reveals different scenarios depending on the criteria but preferred species are indicative of superiority of some of them. The highly ranked based on increasing nutritive value are: (*Maerua Angolensis> Zizyphus mucronata > Acacia Senegal> Acacia mellifera >Balanites aegyptiaca*) but palatability showed a different order: *A. tortilis> M. angolensis> B. aegyptiaca> Z. mucronata Albizia coriaria >* and *Z. mucronata. M. angolensis* and *Z.mucronata* were comparable in intake (358 to 638 gDM/day) and rumen NH₃-N (9.3-13.7mg100ml⁻¹, and ADG (gd⁻¹) between 9.4 to 39.8 g/d. The nutrient digestibility, NH₃-N and ADGs improved with supplementation. It is concluded that *M. angolensis* and *Z. mucronata* can be supplemented alone to improve weight gains of ruminants.

Keywords: Animal performance, antinutritive factors, average daily gains, multipurpose tree forages, rumen degradation, methane amelioration, nutritive values, supplementation

Introduction

In the arid and semi-arid (ASALs) of Sub-Saharan Africa, availability of good quality feed resources all year round is a major constraint to animal production. Animals are offered natural pasture, crop residues or grown fodder such as Napier grass, but most of these feeds are low in digestible nutrients, and when fed alone do not supply adequate nutrients to meet the requirements of the animals ^[50]. The productivity of such animals in terms of meat, milk and other products is, therefore, sub-optimal.

Use of commercial concentrate supplements is one way of improving roughage utilization. However, for smallholder livestock farmers, these concentrates are considered to be too expensive or are not readily available. An alternative to these concentrates is to use on-farm supplements, and forages from multipurpose trees and shrubs (MPTS) ^[25]. The introduced MPTS species such as *Leucaena leucocephala* and *Gliricidia sepium* which are relatively new to the tropics are highly productive and used extensively in areas of high agricultural potential as livestock feed. Nevertheless, they are not adapted to the arid and semi-arid lands where the larger population of livestock is found ^[1, 25, 4, 25, 29]. The native species, like the Acacias, have adapted to the local conditions and are, therefore, better suited for forage production. Besides supplying animals with forages, the tree legumes enrich the soil through biological nitrogen fixation, prevent soil erosion, serve as windbreaks and act as sources of fuel wood and building material, among others.

The MPTS have a wide variation in nutritive value and antinutritive factors. It is, therefore, essential to screen these MPTS using digestibility techniques like the *in sacco* or nylon bag and *in vitro* methods. Some of the antinutritive factors like tannins which abound in tree species like the Acacias ^[113], may compromise the nutritive value of the forages ^[99]. The proanthocyanidins, generally termed tannins, precipitate protein by forming tannin-protein complexes ^[88, 87], thereby reducing the digestibility of protein and feed dry matter ^[33]. The tree forages contain condensed tannins that are very effective in forming tannin-protein complexes. Most studies on MPTS have been on the agronomic performance, identification, propagation

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Corresponding Author: Ondiek JO Egerton University, P.O. Box 536-20115 Egerton, Kenya and management ^[114]. To some extent the basic chemical composition (dry matter, ash, and nitrogen) and feeding value ^[100] have been reported. More work is needed in determining the nutritional role of MPTS in livestock production.

Nutrient Composition of Tree Forages

The nutrient content of forages differs widely among species. Differences may also arise as they grow on different soil types that are endowed with different soil minerals and the forage species have different capacities to manufacture and store the nutrient compounds. It has been shown that tree forages contain higher levels of nutrients than grasses especially in the dry season when the latter wither away. The trees are deep rooted, often tap into the ground water and nutrients and remain green throughout the year ^[3, 22, 64, 72, 73, 74, 78, 93, 103].

Tree forages contain high levels of crude protein (14.7-22.5%) and, therefore, are potential sources of protein supplements. These are higher when compared to grasses like Rhodes Grass (Chloris gayana)^[74] and Maize (Zea mays) stover ^[72] that has been shown to contain 3.6-4.8% and 10% CP, respectively especially in the dry season ^[48]. Leguminous trees that grow easily with minimal agronomic inputs are increasingly becoming alternatives to commercial concentrates. The MPTS are well adapted to the local conditions like soil fertility status, drought regimes, and retain substantial biomass in the dry season apart from withstanding frequent harvesting management [72, 114] and can be harvested and as supplements later [93]. They are moderate to high in minerals that are required by rumen microbes for growth, metabolism and synthetic processes. Low concentration of sulphur and phosphorus in forages reduce fibre digestion ^[97] and minerals in consumed forages in ruminants is scanty and is a growing research area.

Table 1 presents some values for the proximate composition ^[7] of common tree forages browsed by ruminants in the ASALs of Kenya. The values not only vary greatly between and within species but also indicate the ability to supply the much-needed nutrients that may not be the case for grasses.

Table 1: Proximate composition of common tree forages used in						
ASALs of Kenya						

				2	
Species	OM	СР	NDF	ADF	References
A. abyssinica	937	165	462	531	[7, 73, 108, 114]
A. amara	953	167	413	601	[73]; [108]
A. brevispica	927	187	329	460	[1, 4, 7];
A. coriaria	935	169	373	482	[6, 73]
A. elatior	878	162	355	503	[73, 108]
A. hockii	952	121	160	218	[73, 108]
A. mellifera	837	183	306	392	[73]; [103, 104],
A. nilotica	935	121	212	290	[73, 108]
A. senegal	904	249	266	423	[1[4, 16];
A. tortilis	924	117	335	443	[7, 73, 108]
Acacia tortilis pods	-	154	617	-	[103, 104]
B. aegyptiaca	867	137	266	349	[73, 108];
B. aegyptiaca	-	126	283	143	[79, 108]
B. micrantha	940	112	421	481	[73, 108]
Cordia sinensis	-	193	589	568	[103, 108]
Ficus spp	861	412	-	-	[64]
G. bicolor	919	196	362	528	[43, 73, 93, 103, 104]
Justicia exigua	-	204	436	427	[103, 104]
Lannea schweinfurthii	-	174	472	362	[103, 104]
M. angolensis	941	321	332	449	[73]; [78]
Rhus natalensis	-	161	612	308	[103, 104]
Salvadora Persica	-	151	313	239	[103, 104]
Z. mucronata	929	200	222	393	[73]

The Rumen Environment and Feed Utilisation

The rumen environment describes the conditions that affect the breakdown of food in the rumen. These include pH, ammonia level, and a mixture of rumen microflora, which in turn is affected by the former two. Establishing an efficient rumen microbial ecosystem that maximizes fibre digestion and microbial protein synthesis may increase forage utilization by ruminants. For this to occur, sources of readily available and fermentable nitrogen and energy are necessary ^[25, 44]. The rumen ammonia-nitrogen (NH₃-N) is important for efficient synthesis of microbial protein. At low ammonia levels (<5 mg/100 ml) in the rumen fluid, the rumen microflora requires more energy in terms of ATP to fix ammonia into amino acids. Supplementing with fermentable protein sources increases the ammonia levels in the rumen ^{[23,} ^{62]}. The ammonia released by microbial breakdown of protein in the rumen is absorbed through the rumen epithelium and transported in the blood to the liver where it is converted to urea or used by the microbes to synthesize microbial protein and any excess is recycled through saliva and rumen epithelium ^[49, 59] an indication of the adequacy for microbial protein synthesis. The concentration of ammonia in the rumen fluid is influenced by availability of fermentable organic matter or energy and nitrogen, which leads to synchronization of these nutrients to create a higher efficiency of microbial protein synthesis [96]. Supplementing slow energy release grass forage, however, may lead to asynchronous supply of energy and nitrogen in the rumen ^[89] hence low efficiency of microbial protein synthesis.

The minimum rumen NH₃-N levels required for proper rumen function ranges from 5 to 8 mg/100 ml^[92] although levels of 23.5 mg/100 ml may be necessary to maximize feed degradation ^[58, 59, 61]. The rumen NH₃-N range that allow for proper rumen microbial function is wide but the optimum levels for proper microbial function vary from 15-20 mg/100 ml^[60]. The rumen pH normally is in the range between 6 and 7. A pH of 6.2 is the lower limit that allows cellulolysis to proceed properly ^[66]. Therefore, a combination of the right levels of rumen NH₃-N (5-20 mg/100 ml) and pH (6.2-7.0) would optimize feed utilization by animals. The rumen ammonia may be varied depending on the type of feed or supplement given to the animals, but this largely may not influence the methane emission form the animals ^[15] or from the manure of such animals. Other greenhouse gasses (NH₃, N₂O, and CH₄) emission from excreta were not affected as well.

Relative Palatability of Multipurpose Tree Forage

Multipurpose tree and shrub (MPTS) forage has a high potential value as livestock feed and protein supplement ^[48]. The genera *Acacia, Albizia, Leucaena, Gliricidia* and *Prosopis* are among the documented tree species that have forage value. The Acacias predominate in the tropics ^[102, 114] and should be exploited as a quality feed resource.

To assess the palatability ranking of these browses, direct feeding, observation of preference and measurement of intake in confinement is used ^[40] rather than the cafeteria method, which gives an observation on the selection only. ^[39, 40, 45] suggested that good palatability data could be obtained when sheep were adjusted for five days and the feeds provided randomly to control the associative effects developed by animals. It has been reported that the form in which the forage is offered, either wilted or dry did not affect the palatability ranking.

Effect of Tannins on Browse Utilization

Tree forages contain varying levels of phenolic compounds that may contribute to differences in feed utilization by animals. There is usually a reduced intake of feed ^[20] which then affects weight gains due to tannins though low levels of less than 3-6% ^[60] may be beneficial ^[9] and detrimental effects ^[101, 19] set in when this is exceeded. The reactions as well as effects on digestion of those tannins differ with plant species. It is expected that the higher the tannin content in the forage the lower the digestibility and/ or degradability of the material. However, ^[9] found that although Gliricidia contained more tannin than sesbania, the former showed higher rumen degradation.

^[95, 111] demonstrated an increase in dry matter digestibility when gliricidia and leucaena were used as supplements for sheep. When Guinea grass was supplemented with gliricidia in goats, the digestibility of the grass was not affected by supplementation ^[9] though that of nitrogen increased. ^[74] found that gliricidia supplement to Rhodes grass hay for dairy goats increased the total dry matter intake, as found by ^[9] where gliricidia was offered as a supplement to guinea grass diets. ^[3] showed that supplementation of Rhodes grass hay with *Gliricidia sepium* or *Leucaena leucocephala* increased total dry matter intake, and stover intake at lower level of supplementation and this was in contrast to ^[107]. This is comparable with other studies ^[32, 82].

The tannins complex with protein, prevent excessive degradation of protein in the rumen and, therefore, increase the supply of undegradable dietary protein to the lower gut (McNeill *et al*, 1998). This is especially so when the levels of the condensed tannins are moderate, at levels of less than 3-6% ^[60, 64] and may actually reduce digestibility and increase nitrogen excretion in the feces ^[60]. The tannins can be ameliorated by adding polyethylene glycol (PEG) to neutralize their effects ^{[43, 60, 12]; [112]}. Although the protein complexing effect of condensed tannins reduces protein digestibility, it has the advantage of increasing bypass protein by protecting the protein from degradation in the rumen, which becomes available postruminally to the host animal ^[10, 56]. Some browsing animals that feed on browse high in tannins also tend to have high levels of proline in their saliva essentially to neutralize some of the condensed tannins.

Effects of tannins on rumen methane production

The levels of tannins in most tropical multipurpose trees and shrub (MPTS) browses range from 2.9% to 19.7% [26, 98]. These MPTS include species like Leucaena leucocephala, sepium, Gliricidia Samanea saman, Mimosa caesalpiniaefolia, Styzolobium aterrimum, Acacia nilotica and *Acacia mearnsii* ^[14] that have been shown to reduce rumen methane production. ^{[110, 38] and [17]} report on reduction of ruminal enteric methane from ruminants due to inclusion of tannin-rich browses due to the anti-methanogenic activities of tannins. Tropical browses are rich in the tannins that may affect the rumen fauna variably and as a result reduce the population and activity and thus reduce methane production by 3 to 61% [14]. Therefore, an inclusion of some of these

MPTS in ruminant feeds will greatly contribute to reducing anthropogenic methane pollution and general global warming associated with ruminant feeding.

Rumen Degradation and In-Vitro Gas Production

The *in-situ* forage degradation technique is extensively used to evaluate forage quality, but there are inherent material loses due to leakage of fine particles through the pores in the bags. Therefore, pore size and particle size become important factors to consider in this technique. However, mathematical models have been developed to combine degradation and outflow rates that better predict protein degradation ^[76]. This works better for fibrous forages than for concentrate feeds.

Nitrogen deficient feedstuffs like cereal straws and stover form the bulk of basal diet for ruminants in the tropics in the dry season. Intake and digestibility of these feedstuffs is low and animals fed on such materials have reduced production performance. The gas production technique ^[63] simulates the rumen environment and is easily used to predict food digestibility. The rumen microflora requires nutrients for growth and proper function. Nitrogen is one critical nutrient, and 80mg N/L of rumen fluid is required for maximum gas production and carbohydrate degradation. The nitrogen availability affects only the rate and not the extent of degradation. The amount of gas and volatile fatty acids (VFA) produced is not affected by the source of rumen inoculum or the diet consumed by the donor animal if the rumen fluid is taken before feeding [36]. [31] has reviewed the potential of using the gas technique for feed evaluation and outlines the requirements and need to complement it with residue determination.

Effects of Tree Legume Browse on Animal Performance

Supplementing grass diets containing <7% crude protein with legume forages improves both feed intake and animal performance ^[65]. Consequently, there is need to exploit the abundant tree/shrub forage resources. The MPTS forage is a cheap source of nitrogen ^[3] and may lead to significantly higher body weight gains as well as more feed intake when supplemented ^[93] and may form more than 90% of the animals' diet [94]. Acacia pods and leaf litter may sustain sheep, goats and cattle in the dry season, which shows the importance of tree browse as feed for livestock to maximize live weight gains or prevent weight loss, especially when supplementation is provided at 1.0-1.5% of liveweight or 40-60% of DMI ^[70] and using poor quality grass diets ^[67], Stewart et al, 1996, ^{[3, 41]. [3]} concluded that levels of 20 - 30 % of tree legume supplementation could support milk production. The total DMI would increase if energy is also supplemented with the MPTs ^[74, 75]. Some of the highly taniferous species like A. saligna and A. salicinia may negatively affect the animal performance [101, 19].

Table 2 shows the feed intake, weight gain, rumen ammonia nitrogen and nutrient digestibility of Chloris gayana grass supplemented with Maerua angolensis and Zyziphus mucronata mixed browes. Table 2: Dry matter feed intake, average daily gains, rumen ammonia nitrogen and apparent nutrient digestibility of Small East African goatsfed Chloris gayana hay and supplemented with1:1 mixture of Maerua angolensis and Zizyphus mucronata. The treatments studied inclused thecontrol diet that did not have the tree tree forage ad the treatments that had a mixture of M. angolensis and Z, mucrionata at different rates of 15,20, 25 and 30%

Treatments	MZ0	MZ15	MZ20	MZ25	MZ30					
Supplement level, g kg ⁻¹ W ^{0.75}	0	15	20	25	30	SEM				
Dry matter feed intake, gd ⁻¹										
Нау	406 ^a	422 ^a	468 ^b	421 ^a	426 ^a	6.3				
1:1 M. angolensis: Z. mucronata	0.00 ^a	111 ^b	149°	184 ^d	227 ^e	17.8				
Total	406 ^a	533 ^b	617 ^{cd}	605°	653 ^d	20.9				
Total DM intake, % BW	3.1ª	3.9 ^b	4.3°	4.3°	4.7°	0.1				
Daily gain, gd ⁻¹	-4.9 ^a	12.9 ^b	28.1°	14.3 ^b	17.4 ^b	2.7				
Rumen NH ₃ -N, mg/100ml	9.0ª	11.6 ^b	12.7°	11.4 ^b	11.3 ^b	0.3				
Rumen pH	6.9ª	7.0 ^a	6.9 ^a	6.9 ^a	7.0 ^a	0.02				
Арра	rent digestibility	y, g/kg ⁻¹ DM:								
Dry matter	677 ^a	892 ^b	910 ^b	923 ^b	930 ^b	2.3				
Crude protein	627 ^a	716 ^b	758 ^{bc}	793°	767 ^{bc}	1.6				
Organic matter	687 ^a	904 ^b	921 ^b	932 ^b	939 ^b	2.3				
Acid detergent fiber	600 ^a	805 ^b	830 ^b	851 ^b	860 ^b	2.3				
Neutral detergent fiber	506 ^a	847 ^b	875 ^b	892 ^b	902 ^b	3.6				

^{a, b, c} Means on the same row with different superscripts are different at P<0.05 SEM- Standard error of the mean

Source: ^[73].

The Animal Breed Factor and Feed Utilisation

The utilization of tree forages is low despite the high nutrient levels. Due to the high fibre content, their use may require addition of an energy source if they are to contribute fully to the nutritional needs of animals. Energy sources such as cassava peels, molasses or maize bran have been used. Animal breed differences have also been reported ^[35].

Forage utilization by different breeds of ruminant species may vary due to the animal genetic effects on the anatomy and physiology of the animals. This is due to selective breeding that has occurred over time, resulting in sheep flocks suited for local food and environmental conditions. Apart from the chemical and physical factors that influence food digestibility and utilization, animal constraints also play a significant role. The mean retention time of forage in the fore stomach of an animal is related to the reticulo-rumen capacity, level of intake and digestibility of the diet. Rumen size is an anatomical function and, thus a product of the animal's genetics. Rumen fractional clearance rates are generally higher in smaller animals and fast outflow rates have a particular influence on the rumen environment, degradability of fibre and microbial protein yield ^[68, 90]. This hypothesis has led to investigations into differences in the digestive physiology of Bos indicus and Bos taurus cattle, where the former are able to utilize poor quality forage more efficiently. Differences in fermentation rates in the two breeds are reported and that the more tropical, small-sized B. indicus zebu cows degraded fibre better than the more temperate, large-sized cattle [85, 84].

There is likelihood that over time the Small East African Goat breed, adapted to feeding in the marginal areas, has evolved a digestive system that enables it to survive on the poor feed resources. Grasses in these areas are hardy, with a high fibre and low nitrogen levels. Animals kept on such diets are expected to have digestive capacities that are capable of utilizing the grass and shrubs found in these dry lands. This study reports the performance of the local Small East African Goat offered Rhodes grass hay, a low-quality basal diet, supplemented with tree legume leaf forage ^[68, 102].

Factors Determining Feed Intake of Tree Browse

Voluntary feed intake is the weight of feed eaten by an animal or group of animals during a given period of time during which they have free access to food ^[28].

Consumption of food initiates several events important for eventual development of satiety. Absorbed nutrients, gut hormones and sensory nerves are believed to mediate feedback mechanisms. Since animal productivity is largely dependent on the intake of food, the stockman strives to increase the voluntary intake of farm animals. The importance of the quantity of food an animal takes in is emphasized by the fact that the more food an animal consumes each day, the higher is the level of ingested nutrient and, therefore, the greater will be the opportunity for increasing its daily production.

Ruminant intake is controlled by a combination of physical, chemical and psychological factors. Of the four mechanisms of feed regulation, that is glucostatic, chemostatic, thermostatic and lipostatic, the glucostatic mechanism looks the least likely to influence feed intake in ruminants ^[45]. This is because the absorption of glucose in the gastrointestinal tract of ruminants is either absent or very low as most of the sugars are fermented to volatile fatty acids. In the tropics, performance of ruminant livestock is largely limited by voluntary intake and digestibility of the basal feed, which mainly consists of low-quality roughages and crop residues ^[45, 82]. There are five main factors that determine feed intake of tree browses, including animal factors [34, 109], environmental factors [109], physical factors [83], chemical factors [8, 32, 52, 73, 74, 82, 91] and antinutritional factors in tree legume browse ^[2, 3, 6, 11, 12, 18, 19, 22, 47, 54, 57, 72, 86, 88, 101, 106, 113]

Antinutritional Factors in Tree Legume Browse

Treating forages with chemicals such as polyethylglycol (PEG) or polyvinyl pyrridone (PVP) reduces the tannin content of the feed and the corresponding anti-nutritive effect. In a study designed to investigate the effect of administering PEG on tannin-rich diets significant increases in DMI were noted when PEG was added to diets of *A. saligna* and those of quebracho powder (tannic acid) ^[20]. Spraying high-tannin containing *Lotus pedunculatus* with PEG reduced the total reactive tannin content from 63 to 7 g/kg DM. This resulted in an increase in the voluntary intake of digestible organic matter (DOM), digestible fibre (P<0.05), and digestive total N (P<0.01) ^{[12]; [112]} with the lower intakes associated with

negative effects on rumen fermentation rather than palatability.

The inclusion of tree legume browse in grass and crop residue ruminant diets is expected to improve performance, be it growth, milk or wool production. However, the presence of ANFs may be an impediment to expected performance unless their effects are minimized or their inhibitive effects are countered by suitable nutrient supplementation. Negative effects on production due to the ANFs could be countered by the additional provision of urea. The synergistic effect of combining rumen supplements with by-pass nutrients leads to dramatic improvement in animal performance, especially for demanding functions such as milk production. The urea will be an additional supply of N directly to the animal. The presence of the ANFs, especially phenolic compounds in tree legumes may generally enhance their usefulness as supplements.

Geographic spread and abundance

^[73] assessed fifteen indigenous Kenyan browse species including *Maerua angolensis, Acacia brevispica* (Wait-a-bit thorn), *Acacia mellifera* (Oiti, Maasai), *Acacia tortilis, Acacia hockii* (Enchapalani, Maasai), *Zizyphus mucronata* (Buffalo Thorn, Cape thorn), *Grewia bicolor* (Sitetet, Kipsigis), *Acacia elatior* (Olerai, Maasai), *Acacia nilotica* (Nile thorn) *Balanites aegyptiaca* (Desert date), *Acacia senegal* (Gum Arabic Acacia, Gum Arabic tree), *Acacia abyssinica* (Flat-top Acacia), *Bridelia micrantha* (Shikanganya, Luhya), *Albizia amara* (Bitter albizia), and *Albizia coriaria*.These are widespraed in the Kenyan ASALs and are favourable due to not only their moderate to high nutritive value and were used for animal performance experiments but alsothe species abundance, robustness and ability to withstand dry conditions [⁸¹] and versatile uses as well ^{[48]; [114]}.

The true nutritional values of each species is rated on the crude protein content (112-321gkg⁻¹DM) of the species where ^[21] reported similar values ranging of 10.5 to 22.5% DM and these are high compared to pasture grasses (48-52gkg⁻¹ DM). They contain relatively high macro and micro mineral content, especially calcium and iron ^[2, 29, 30, 48]. The species are considerably rich in most of the minerals: Ca, P, Mg, Fe, Mn, Cu, Mo Zn and Se and, therefore, can be good sources of these mineral ^[2, 42, 93]. The NDF content of 218 to 601gkg⁻¹ DM and ADF of 160 to 462gkg⁻¹DM is good as the higher the ADF and lower NDF fractions in feeds, the higher the digestible organic matter and expected metabolisable energy value. However, there could be a negative correlation between palatability and NDF, ADF and lignin, and a positive correlation to NDF-N ^[27, 39, 70, 71]. The palatability could be influenced by the polyphenol or tannin contents ^[73] that may affect the organic matter digestibility [47, 54, 86].

The tannins levels have beneficial effects, mediated by protein-tannin complexation at rumen pH and the dissociation of the complex post-ruminally, enhancing availability of feed protein for production purposes ^[10] although this depends on the nature of tannins and its affinity towards macromolecules ^[53]. Tannins at low levels could help synchronize the release of various nutrients, which in turn might be responsible for increase in microbial efficiency by increasing supply of non-ammonia nitrogen in the lower intestine for production ^[53].

The different palatability indices used in the palatability studies did not show significant differences in the final results. The procedure by ^[48] could be recommended because

the palatability index is based on the DM intakes of both the test forage and the basal diet. This method is preferable because the quantity and quality of the basal diet can influence the intake of a supplement ^[76]. The number of days used did not also show any significant change, though ^[13] recommend a recording of the intakes after an adaptation period of at least one day, and observations for 5 to 12 days.

Supplementation has been defined as the addition on a daily basis of a proportion of a forage to a basal diet, with the supplementary forage obtained either by grazing, as cut forage or a by-product from another industry ^[24]. The leaves of *A. brevispica* leaves have been shown to be as good as Lucerne hay and *A. tortilis* pods when given to sheep offered a basal diet of poor-quality grass hay ^[37]. Supplementation of barley straw diet for wethers with *A. brevispica* leaves and urea greatly improved the value of the overall diet. This was observed in the resulting improved intakes, digestibilities and weight gains. The increase in the feed intakes may be due to the species' relatively higher N-content than grasses as demonstrated by ^[80, 105, 82]. However, higher nitrogen excretion may lower animal performance despite having the high N intake ^[69]. This can be improved by supplying additional energy to enhance nutrient ^[74, 75, 96].

Conclusions

The following conclusions can be drawn from this review:

- 1. The ranking of tree forages based on nutrient composition, particularly CP, minerals, *in-vitro* and *in-vivo* degradation and 48hr OMD reveals the best forages that are potential supplements for poor quality basal feeds.
- Multipurpose tree fodders are palatable to goats, especially *A. tortilis, M. angolensis, B. aegyptiaca, Z. mucronata, A. coriaria, A. Senegal* and *A. abyssinaica* which have a palatability index range of 1.8-1.2 and *Z. mucronata* and *M. angolensis* are protein-rich (200gkg⁻¹ DM and 321 gkg⁻¹ DM) with positive diet intake, digestibility and average daily gains.

Implications

Leaf forages from the local MPTS can be used as protein supplements and mineral sources to ameliorate these nutrient deficiencies for ruminant livestock feeding on poor basal diets and the leaf forages can be harvested and stored to be fed with basal diets especially in the dry season of feed scarcity.

References

- 1. Abdulrazak SA, Nyangaga J, Fujihara T. Relative Palatability to Sheep of Some Browse Species Their In scco Degradability and *in vitro* Gas Production Characteristics - Abdulrazak 2001; c2001.
- 2. Abdulrazak SA, Fujihara T, Ondiek JO, Ørskov ER. Nutritive evaluation of some Acacia tree leaves from Kenya. Anim Feed Sci Technol. 2000;85:89-98.
- 3. Abdulrazak SA, Muinga RW, Thorpe W, Ørskov ER. The effect of incremental supplementation with Gliricidia sepium and Leucaena leucocephala on voluntary food intake, rumen fermentation, microbial N supply and liveweight changes of *Bos taurus* x *Bos indicus* steers offered Zea mays stover ad libitum. Livest Prod Sci. 1997;49:53-62.
- 4. Abdulrazak SA, Nyangaga JN, Fujihara T. Relative palatability, in-sacco degradation and *in-vitro* gas production of some multipurpose fodder trees. Asian-Australas J Anim Sci. 2001;14(11):1580-1586.

- 5. Adjorlolo LK, Amaning-Kwarteng K, Fianu FK. *In vivo* digestibility and effect of supplemental *Mucuna* forage on treated rice straw degradation. Small Rumin Res. 2001;41:239-245.
- Adugna T, Khazaal K, Ørskov ER. Nutritional evaluation of some browse species. Anim Feed Sci Technol. 1997;67:181-195.
- AOAC. Association of Official Analytical Chemists. Official Methods of Analysis. Washington, DC: AOAC; 1990.
- 8. ARC. The nutrient requirement of ruminant livestock. Technical review by an agricultural research council working party. Published on behalf of ARC by CAB International; c1980.
- 9. Ash AJ. The effect of supplementation with leaves from leguminous trees *Sesbania grandiflora*, *Albizia chinensis* and *Gliricidia sepium* on the intake and digestibility of Guinea grass hay by goats. Anim Feed Sci Technol. 1990;28:225-231.
- Barry TN, Manly TR, Duncan SJ. The role of condensed tannins in the nutritional value of *Lotus pendanculatus* for sheep, 4. Sites of carbohydrate and protein digestion as influenced by dietary reactive tannin concentration. Br J Nutr. 1986;55:123-137.
- Ben Salem H, Nefzaoui A. Effect of increasing level of Acacia cyanophylla Lindl. on intake, digestibility and ruminal digestion in sheep fed straw-based diets. In: Nikolaidis A, Papanastis V, editors. Management of Mediterranean shrublands and related forage resources. Rome: FAO; 1993. p. 118-121.
- Ben Salem H, Nefzaoui A, Abouli H. Palatability of shrubs and fodder trees measured on sheep and dromedaries: 1. Methodology approach. Anim Feed Sci Technol. 1994;46:143-153.
- 13. Ben Salem H, Nefzaoui A, Ben Salem L, Tisserand JL. Effect of *Acacia cyanophylla* Linndl. Forage supply on intake and digestion by sheep fed Lucerne hay based diets. Anim Feed Sci Technol. 1997;68:101-113.
- Cardoso-Gutierrez E, Aranda-Aguirre E, Robles-Jimenez LE, Castelan-Ortega OA, Chay-Canul AJ, Foggi G, *et al.* Effect of tannins from tropical plants on methane production from ruminants: A systematic review. Vet Anim Sci. 2021;14:100214.
- Carvalho GM, Brito LF, Coelho LdeF, Melo C, Cardoso AdS, Messana JD, *et al.* Agronomy Journal. 2022;114(3):1571-1893.
- Chen XB, Deb Hovell F, Dube JS, Reed JD, Ndlovu LR. Proanthocyanidins and other phenolics in Acacia leaves of Southern Africa. Anim Feed Sci Technol. 2001;91:59-67.
- 17. Costa M, Alves SP, Cabo A, Guerreiro O, Stilwell G, Dentinho MT, *et al.* Modulation of *in vitro* rumen bio hydrogenation by *Cistus ladanifer* tannins compared with other tannin sources. J Sci Food Agric. 2018;97(2):629-635.
- D'Mello JPF. Chemical constraints to the use of tropical legumes in animal nutrition. Anim Feed Sci Technol. 1992;38:237-261.
- 19. Degen AA, Blanke A, Becker K, Kam M, Benjamin RW, Makkar HPS. The nutritive value of *Acacia salicinia* for goats and sheep. Anim Sci. 1997;64:253-259.
- 20. Degen AA, Mishorr T, Makkar HPS, Kam M, Benjamin RW, Becker K, *et al.* Effect of *Acacia saligna* with and without administration of polyethylene glycol on dietary intake in desert sheep. Anim Sci. 1998;67:491-498.

- 21. Devendra C. Small farm systems combining crops and animals. In: Proceedings of the Fifth World Conference on Animal Production. 1983;1:173-191.
- 22. Ebong C. Acacia nilotica, Acacia seyal and Sesbania sesban as supplements to Teff (Eragrostisteff) straw fed to sheep and goats. Small Rumin Res. 1995;18:233-238.
- 23. Egan AR. Principles of supplementation of poor quality roughage with nitrogen. In: Dixon RM, editor. Ruminant Feeding Systems Utilizing Fibrous Agricultural Residues-1985. Canberra: International development Programme of Australian universities and Colleges limited (IDP); c1986. p. 49-57.
- 24. Elliot R, McMeniman MP. Supplement of ruminant diets with forage. In: Hacker JB, Ternouth JH, editors. The Nutrition of Herbivores. Sydney: Academic Press; c1987.
- 25. Elseed FAMA, Amin AE, Khadiga A, Ati A, Sekine J, Hishinuma M, *et al.* Nutritive evaluation of some fodder tree species during the dry season in Central Sudan. Asian-Austral J Anim Sci. 2002;6:844-850.
- 26. Fernandes AP, Shintre PM, Fulpagare YG. Nutritive value of top feeds for goats. Indian J Anim Nutr. 2007;24(1):40-43.
- 27. Fonseca MS, Kenworthy J, Thayer GW. Science for Solutions NOAA'S COASTAL OCEAN PROGRAM Decision Analysis Series No. 12 ines for t e Conservation and Restoration e United States ters. 1998. Available from: http://www.cop.noaa.gov
- 28. Forbes JM. Voluntary food intake and reproduction. Proc Nutr Soc. 1987;46(2):193-201.
- 29. Fujihara T, Hosoda C, Matsui T. Mineral status of grazing sheep in the dry area of mainland China. Asian-Austral J Anim Sci. 1995;8:179-186.
- Fujihara T, Matsui T, Hayashi S, Robles AY. Mineral status of grazing Philippine goats II. The nutrition of selenium, copper and zinc of goats in Luzon Island. Asian-Austral J Anim Sci. 1992;5:389-395.
- Getachew G, Blümmel M, Makkar HPS, Becker K. In vitro gas measuring techniques for assessment of nutritional quality of feeds: A review. Anim Feed Sci Technol. 1998;72:261-281.
- Goodchild AV, Mcmeniman NP. Intake and Digestibility of Low Quality Roughages When Supplemented with Leguminous Browse. J Agric Sci. 1994;122(1):151-160. doi:10.1017/S0021859600065904
- Hangarman AE, Robins CT, Weerasuriya Y, Wilson TC, McArthur C. Tannin chemistry in relation to digestion. J Range Manage. 1992;45:57.
- 34. Hayirli A, Grummer RR, Nordheim EV, Crump PM. Animal and Dietary Factors Affecting Feed Intake During the Prefresh Transition Period in Holsteins. J Dairy Sci. 2002;85(12):3430-3443. doi:10.1002/agj2.21053
- 35. Hungate RA, Phillips GD, Hungate DP, MacGregor A. A comparison of the rumen fermentation in European and Zebu cattle. J Agric Sci. 1960;54:196–201.
- 36. Huntington JA, Rymer C, Givens DI. The effect of host diet on the gas production profile of hay and high-temperature dried grass. Anim Sci. 1998;67:59-64.
- 37. International Livestock Centre for Africa (ILCA). Livestock Development in Sub-Saharan Africa Postdrought Vegetation in the Niger Floodplain Sheep Mortality in Highland, Ethiopia; c1998.
- 38. Jayanegara A, Goel G, Makkar HPS, Becker K. Divergence between purified hydrolysable and condensed tannins effects on methane emission, rumen fermentation

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and microbial population *in vitro*. J Anim Feed Sci Technol. 2015;209:60-68.

doi:10.1016/j.anifeedsci.2015.08.002.

- 39. Kaitho RJ, Umnnaa NN, Nsahlai IV, Tamminga S, Van Bruchem J, Hanson J, *et al.* Palatability of wilted and dried multipurpose tree species fed to sheep and goats. Anim Feed Sci Technol. 1997;65:151-163.
- 40. Kaitho RJ, Umunna NN, Nsahlai IV, Tamminga S, Van Bruchem J, Hanson J, Van de Wouw M. Palatability of multipurpose tree species: Effect of species and length of study on intake and relative palatability by sheep. Agrofor Syst. 1996;33:249-261.
- 41. Kanjanapruthipong J, Leng RA. The effects of dietary urea on microbial populations in the rumen of sheep. Asian-Australas J Agric Sci. 1998;11:661-672.
- 42. Karugia JT, Nyangaga J, Waithaka MM, Mwangi DM, Sinja J, Romney DL. Factors influencing adoption and dissemination of forage legumes in central Kenya. 2004. http://hdl.handle.net/11295/81503
- 43. Kemboi F, Ondiek JO, King'ori AM, Onjoro PA. Effects of incorporation of selected browse species on performance of growing small East African goats. Livest Res Rural Dev. 2021, 33(8).
- 44. Kendall NR, Mackenzie AM, Telfera SB. Effect of a copper, cobalt and selenium soluble glass bolus given to grazing sheep. Livest Prod Sci. 2001;68:31-39.
- 45. Khan LP, Quellet D, Nolan JV. Estimation of allantoin flux using continuous infusion of allantoin sensitivity to plasma loading with unlabelled allantoin. Br J Nutr. 2001;86:691-696.
- 46. Khanal RC, Subba DB. Nutritional evaluation of leaves from major fodder trees cultivated in the hills of Nepal. Anim Feed Sci Technol. 2001;92:17-32.
- 47. Kumar R, Vaithiyanathan S. Occurrence, nutritional significance and effect on animal productivity of tannins in tree leaves. Anim Feed Sci Technol. 1990;30:21-38.
- Le Houerou HN. Indigenous shrubs and trees in silvopastoral systems of Africa. In: Steppler HA, Nair NKR, eds. Agroforestry: A Decade of Development. Nairobi: ICRAF; c1987. p. 139-156.
- Leaver JD. Milk production. Longman handbooks in Agriculture. Astros Printing Ltd. Hong Kong; c1983. p. 61-63.
- 50. Leng RA. Factors affecting the utilization of 'poor quality' forages by ruminants particularly under tropical condition. Nutr Res Rev. 1990;3:277-303.
- Leng RA, Nolan JV. Nitrogen metabolism in the rumen. J Dairy Sci. 1984;67:1072-1089.
- 52. Mahgoub O, Lu CD, Early RJ. Effect of Dietary Energy Density on Feed Intake, Body weight gain and Carcass Chemical Composition of Omani growing lambs. Small Rumin Res. 2000;37:35-42.
- 53. Makkar HPS. Effects and Fate of Tannins in Ruminant Animals, Adaptation to Tannins, and Strategies to Overcome Detrimental Effects of Feeding Tannin-Rich Feeds. Small Rumin Res. 2003;49:241-256. doi:10.1016/S0921-4488(03)00142-1
- Makkar HPS. Antinutritional factors in foods for livestock. In: Animal Production in Developing Countries, Occasional Publication No. 16. BSAP (Eds. M. Gill, E. Owen, G. E. Pollot and T. L. J. Lawrence); c1993. p. 69-81.
- 55. Makkar HPS. Quantification of Tannins in Tree Foliage. A laboratory manual for the FAO/IAEA Co-ordinated Research Project on, "use of nuclear and related

techniques to develop simple tannin assays for predicting and improving the safety and efficiency of feeding ruminants on tanniferous tree foliage". FAO/IAEA working document. IAEA, Vienna, Austria; 2000. 26pp.

- 56. Makkar HPS, Becker K. Adaptation of cattle to tannins: role of proline-rich proteins in oak-fed cattle. Anim Sci. 1998;67:277-281.
- 57. Mangan JL. Nutritional effects of tannins in animal feeds. Nutr Res Rev. 1988;1:209-231.
- McDonald I. A revised model for the estimation of protein degradability in the rumen. J Agric Sci. 1981;96:251-258.
- McDowell LR. Nutrition of grazing ruminants in warm climates. Academic Press/Harcourt Brace, London; c1985.
- 60. McNeill DM, Osborne N, Komolong MK, Nankervis D. Condensed tannins in the genus Leucaena and their nutritional significance for ruminants. In: Leucaena – Adaptation, quality and farming systems. Proceedings of a workshop held in Hanoi, Vietnam 9-14 February, 1998. Australian Centre for International Agricultural Research Proceedings No. 86; c1998. p. 205-214.
- 61. Mehrez AZ, Ørskov ER, McDonald I. Rates of rumen fermentation in relation to ammonia concentration. Br J Nutr. 1977;38:437-443.
- 62. Melaku S, Peters KJ, Tegegne A. *In-vitro* and *in-situ* evaluation of selected multipurpose trees, wheat bran and Lablab purpureus as potential feed supplements to Teff (Eragrostis tef) straw. Anim Feed Sci Technol. 2003;108:159-179.
- 63. Menke KH, Steingass H. Estimation of the energetic feed value obtained from the chemical analysis and *in vitro* gas production using rumen fluid. Anim Res Dev. 1988;28:7-55.
- 64. Minal HP, Malek SS, Patel VR, Desai BS, Jha SK, Patel DP. Chemical composition and tannin content in leaves of Ficus spp. as top feed for ruminants from the Dang forest of South Gujarat Province, India. Int J Vet Sci Anim Husbandry. 2023;8(3):225-228. doi:10.22271/veterinary.2023.v8.i3d.548
- 65. Minson DJ, Milford R. The voluntary intake and digestibility of diets containing different proportions of legume and mature Pongola Grass (*Digitaria decumbens*). Aust J Exp Agric Anim Husb. 1967;7:546-551.
- 66. Mould FL, Ørskov ER. Manipulation of rumen pH and its influence on the cellulolysis, in-sacco dry matter degradation and rumen microflora of sheep offered either hay or concentrate. Anim Feed Sci Technol. 1984;10:1-4.
- 67. Muinga RW, Thorpe W, Topps JH. Voluntary food intake, liveweight changes and lactation performance of cross-bred dairy cows given ad libitum Pennisetum purpureum (*Napier grass* var. Bana) supplemented with Leucaena forage in the lowland Semi-humid tropics. Anim Prod. 1992;55:331-337.
- 68. Nantoume H, Forbes TDA, Hensarling CM, Sieckenius SS. Nutritive value and palatability of guajillo (*Acacia berlandieri*) as a component of goat diets. Small Rumin Res. 2001;40:139-148.
- Nherera FV, Ndlovu LR, Dzowela BH. Utilisation of Leucaena diversifolia, Leucaena esculenta, Leucaena pallida and Calliandra calothyrsus as nitrogen supplements for growing goats fed maize stover. Anim Feed Sci Technol. 1998;74(1):15–28. doi:10.1016/S0377-8401(98)00164-3

- 70. Norton BW. Tree legumes as dietary supplements for ruminants. In: Gutteridge RC, Shelton HM, editors. Forage Tree Legumes in Tropical Agriculture. CAB International; c1994. p. 177-191. Available from: http://www.fao.org/Ag/agp/agpc/doc/PUBLICAT/Guttshel/x5556e0k.htm
- Nsahlai IV, Siaw DEKA, Osuji PO. The relationship between gas production and chemical composition of 23 browses of genus Sesbania. J Sci Food Agric. 1994;65:13-20.
- 72. Odenyo AA, Osuji PO, Karanfil O, Adinew K. Microbiological evaluation of *Acacia angustissima* as a protein supplement to sheep. Anim Feed Sci Technol. 1997;65:99-112.
- 73. Ondiek JO, Abdulrazak SA, Njoka EN. Chemical and mineral composition, *in-vitro* gas production, *in-sacco* degradation of selected indigenous Kenyan browses. Livest Res Rural Dev. 2010;22. Available from: http://www.lrrd.org/lrrd22/2/ondi22025.htm
- 74. Ondiek JO, Abdulrazak SA, Tuitoek JK, Bareeba FB. The effects of *Gliricidia sepium* and maize bran as supplementary feed to Rhodes grass hay on intake, digestion and liveweight of dairy goats. Livest Prod Sci. 1999;61:65-70.
- 75. Ondiek JO, Tuitoek JK, Abdulrazak SA, Bareeba FB, Fujihara T. Use of *Leucaena leucocephala* and *Gliricidia sepium* as nitrogen sources in supplementary concentrates for dairy goats offered Rhodes grass hay. Asian-Australas J Anim Sci. 2000;13:1249-1254.
- 76. Ørskov ER, McDonald I. The estimation of protein degradability in the rumen from incubation measurements weighed according to rate of passage. J Agric Sci. 1979;92:499-503.
- 77. Osuga IM, Wambui CC, Abdulrazak SA, Ichinohe T, Fujihara T. Evaluation of nutritive value and palatability by goats and sheep of selected browse foliages from semiarid area of Kenya. Anim Sci J. 2008;79(5):582– 589. doi:10.1111/j.1740-0929.2008.00567.x
- 78. Osuga IM, Abdulrazak SA, Ichinohe T, Fujihara T. Rumen degradation and *in-vitro* gas production parameters in some browse forages, grasses and maize stover from Kenya. J Food Agric Environ. 2006;4:60-64.
- Ouedraogo P, Kere M, Tianhoun D, Fidèle TA, Kabore A, Tamboura HH, Belem AMG. *Balanites aegyptiaca* (L.) Delile leaves nutritional value for local Mossi sheep in Burkina Faso. Int J Vet Sci Anim Husbandry. 2023;8(3):85-89.

doi:10.22271/veterinary.2023.v8.i3b.527

- Pathirana C, Jensen P, Fenical W. Marinone and debromomarinone: Antibiotic sesquiterpenoid naphthoquinones of a new structure class from a marine bacterium. Tetrahedron Lett. 1993;33(50):7663–7666. doi:10.1016/0040-4039(93)88010-G
- Pellew RA. The giraffe and its food resource in the Serengeti. II. Response of the giraffe population to changes in the food supply. J Ecol. 1983 Dec;21(4):269-283.
- 82. Premaratne S, Bruchem VJ, Chen XB, Perera HGD, Oosting SG. Effects of type and level of forage supplementation on voluntary intake, rumen microbial synthesis and growth in sheep fed a basal diet of rice straw and cassava. Asian-Australas J Anim Sci. 1998;11:692-696.

- 83. Ramirez RG, Ledezma-Tores RA. Forage utilization from native shrubs *Acacia regula* and *Acacia fernesiana* by goats and sheep. Small Rumin Res. 1997;25:43-50.
- Ranilla M, Lopez S, Giraldez FJ, Valdes C, Carro MD. Comparative digestibility and digesta flow kinetics in two breeds of sheep. J Anim Sci. 1998;66:389-396.
- 85. Ranilla MJ, Carro MD, Valdes C, Giraldez FJ, Lopez S. A comparative study of ruminal activity in Churra and Merino sheep fed alfalfa hay. Anim Sci. 1997;65:121-128.
- Reed JD. Relationship among soluble phenolics, insoluble phenolics, insoluble proanthocyanidins and fibre in East African browse species. J Range Manage. 1986;39:5-6.
- 87. Reed JD. Nutritional toxicology of tannins and related polyphenols in forage legumes. J Anim Sci. 1995;73:1516-1528. Available from: http://jas.fass.org/cgi/reprint/73/5/1516.pdf
- 88. Reed JD, Soller H, Woodward A. Fodder tree and straw diets for sheep: Intake, growth, digestibility and the effects of phenolics on nitrogen utilisation. Anim Feed Sci Technol. 1990;30:39-50.
- Richards DE, Brown WF, Ruegsegger G, Bates DB. Replacement value of tree legumes for concentrates in fodder-based diets. I. Replacement value of Gliricidia sepium for growing goats. Anim Feed Sci Technol. 1994;46:37-51.
- Salawu MB, Acamovic T, Stewart CS, Roothaert RL. Composition and degradability of different fractions of Calliandra leaves, pods and seeds. Anim Feed Sci Technol. 1999;77:181-199.
- 91. Sarwar M, Mahr-un-Nisa, Bhatti SA, Ali CS. In situ Ruminal Digestion Kinetics of Forages and Feed Byproducts in Cattle and Buffalo. Asian-Australas J Anim Sci. 1998;11(2):128–132. doi:10.5713/ajas.1998.128
- 92. Satter LD, Slyter LL. The effect of ammonia concentration on rumen microbial protein production *in vitro*. Br J Nutr. 1974;32:194-208.
- 93. Sawe JJ, Tuitoek JK, Ottaro JM. Evaluation of common tree leaves or pods as supplements for goats on range area of Kenya. Small Rumin Res. 1998;28:31-37.
- 94. Scholte PT. Leaf litter and Acacia pods as feed for livestock during the dry season in *Acacia* –Commiphora bushland, Kenya. J Arid Environ. 1992;22:271-276.
- 95. Serra AB, Serra SD, Serra FB, Domingo IJ, Cruz LC, Fujihara T. Diets of the Philippine indigenous sheep: Its comparison to indigenous goats diets and influence of sampling methods. Asian-Australas J Anim Sci. 1995;8:163-169.
- 96. Sinclair LA, Garnsworthy PC, Newbold JR, Buttery PJ. The effect of synchronising the rate of dietary energy and nitrogen release in diets with similar carbohydrate composition and rumen fermentation and microbial protein synthesis in sheep. J Agric Sci. 1995;120:251-263.
- 97. Spears JW. Forage quality evaluation and utilisation. In: Fahey GC Jr, Mertens DR, Collins M, Moser LE, editors. National Conference on forage quality evaluation and utilisation, 1994. Held at the University of Nebraska; c1995. p. 281-317.
- 98. Sumi SA, Siraj MA, Hossain A, Mia MS, Afrin S, Rahman MM. Pharmacological activities of Ficus racemosa and analysis of its major bioactive polyphenols

by HPLC-DAD. Highlights on Medicine and Medical Science. 2021;5:68-84.

- 99. Swain T. Tannins and lignins. In: Rosenthal G, Jansen DH, editors. Herbivores, their interaction with secondary plant metabolites. Academic Press; c1979. p. 657.
- 100. Tamang BB, Shah MK, Dhakal B, Chaudhary P, Chhetri N. Participatory ranking of fodders in the western hills of Nepal. J Agric Nat Resour. 2020;3(1):20-28. doi:https://doi.org/10.3126/janr.v3i1.27001
- 101. Tanner JC, Reed JD, Owen E. The nutritive value of fruits (pods wit seeds) from four Acacia species compared to extracted noug (*Guizotia abyssinica*) meal as supplement to maize stover for Ethiopian Highland sheep. Anim Prod. 1990;51:127-133.
- 102. Topps JH. Potential, composition and use of legume shrubs and trees as fodder for livestock in the tropics (a review). J Agric Sci. 1992;118:1-8.
- 103.Tura I, Ondiek J, King'ori A, Onjoro P. Economic Analysis of Feeding Locally Formulated and Commercial Milk Replacer to Camel Calves in the Arid and Semi-Arid Lands of Kenya. Arch Bus Res. 2022;10(8):207-223.
- 104. Tura I, Ondiek JO, Kingo'ri AM, Onjoro PA. Proximate composition of selected browses and common milk supplements for camel calves in Kenya. Int J Vet Sci Anim Husb. 2021;6(5):31-39.
- 105.Umunna NN, Nsahlai IV, Osuji PO. Degradability of forage protein supplements and their effects on the kinetics of digestion and passage. Int Livestock Centre for Africa (ILCA); c1995.
- 106. Vaithiyanathan S. Salivary gland tannins binding proteins of sheep and goat. Indian J Anim Sci. 2001; 71(12) 1131-1134.

Available from:

https://www.researchgate.net/publication/286960747

- 107. Van Eys JE, Mathius IE, Pongsapan P, Johnson WL. Foliage of tree legumes *Gliricidia*, *Leucaena*, and *Sesbania* as supplement to napier grass diets for growing goats. J Agric Sci. 1986;107:227-233.
- 108.Van Soest PJ, Robertson JD, Lewis BA. Methods of dietary fiber, neutral detergent fiber and non-starch polysaccharides in relation to animal nutrition. J Dairy Sci. 1991;74:3583-3597. Available from: http://jds.fass.org/cgi/reprint/74/10/3583.pdf
- 109.Van DTT. Some Animal and Feed Factors Affecting Feed Intake, Behaviour and Performance of Small Ruminants. Doctoral thesis Swedish University of Agricultural Sciences Uppsala; c2006.
- 110. Vasta V, Daghio M, Cappucci A, Buccioni A, Serra A, Viti C, *et al.* Invited review: plant polyphenols and rumen microbiota responsible for fatty acid biohydrogenation, fiber digestion, and methane emission: experimental evidence and methodological approaches. J Dairy Sci. 2019;102(5):3781–3804. doi:10.3168/jds.2018-14985
- 111.Veereswara BR, Parthasarathy M, Krishna N. Effect of supplementation with tree leaves on intake and digestibility of hybrid Napier (NB-21) grass in Nellore sheep. Anim Feed Sci Technol. 1993;44:265-274.
- 112. Waghorn GC, Shetton ID, McNabb WC, McCutcheon. Effects of condensed tannins in *Lotus pedunculatus* on its nutritive value for sheep, 2. Nitrogenous aspects. J Agric Sci. 1994;123:109-117.
- 113.Woodward A, Reed JD. Nitrogen metabolism of sheep and goats consuming *Acacia brevispica* and *Sesbania*

sesban. J Anim Sci. 1997;75:1130-1139. Available from: http://jas.fass.org/cgi/reprint/75/4/1130.pdf

114.World Agroforestry Centre (WAC). Useful trees and shrubs for Kenya. World Agroforestry Centre –Eastern and Central Africa Regional Programme (ICRAF, 2005-ECA). Technical Handbook No. 35. Eds. Patrick Maundu and Bo Tengnas. Nairobi, Kenya; c2005. p. 484.