



ISSN: 2456-2912

VET 2024; 9(1): 174-178

© 2024 VET

www.veterinarypaper.com

Received: 15-11-2023

Accepted: 25-12-2023

Manoj Kumar Kalwaniya

Department of Veterinary Public Health and Epidemiology, College of Veterinary and Animal Science, Navania Vallabh Nagar, Udaipur, Rajasthan, India

Devender Choudhary

Department of Veterinary Public Health and Epidemiology, College of Veterinary and Animal Science, Navania Vallabh Nagar, Udaipur, Rajasthan, India

Surendra

Department of Veterinary Public Health and Epidemiology, College of Veterinary and Animal Science, Navania Vallabh Nagar, Udaipur, Rajasthan, India

Corresponding Author:

Manoj Kumar Kalwaniya

Department of Veterinary Public Health and Epidemiology, College of Veterinary and Animal Science, Navania Vallabh Nagar, Udaipur, Rajasthan, India

***Escherichia coli* prevalence and antibiograms identified in meat or meat products**

Manoj Kumar Kalwaniya, Devender Choudhary and Surendra

Abstract

An essential pathogen that is found in food is *Escherichia coli*. It serves as a facultative anaerobic bacterium that is harmful to both humans and animals and is part of normal microbial flora of digestive tract. Diarrheagenic *E. coli* are those that cause diarrhea along with other enteric illnesses. Although many strains of *E. coli* colonize the intestine innocuously, others cause a variety of intestinal illnesses. *E. coli* contamination of meat as well as meat products poses a risk to public health. 100 samples of RTE meat products (30), chevon (20), mutton (20), and chicken meat (30) were gathered from the Rajasthani city of Udaipur. The chicken, mutton, chevon, and RTE meat products all had *E. coli* incidence levels of 56.66% (17), 45% (9), 40% (8), and 13.33% (4), respectively. Antibiotic susceptibility testing against ten different antibiotics were also performed on all *E. coli* isolates of meat/meat products. The results of the antibiogram study showed that the isolates were susceptible to ceftriaxone (78.95%), gentamicin (78.95%), and ampicillin (63.16%). While erythromycin (94.74%), Amoxicillin+Clavulanic acid (68.4%), and Oxytetracycline (68.4%) were all shown to be resistant to the isolates.

Keywords: Meat products, *E. coli*, prevalence, antibiogram

Introduction

Worldwide public health challenges include microbial food safety and food-borne illnesses [9]. The high biological value, low cost, wonderful flavor, and ease of preparation, meat and or ready-to-eat (RTE) meat products are in highly demanded [32, 39]. They are also excellent source of good quality proteins, mineral, and vitamin. The World Health Organization claims that a significant part of diarrhea around the world is brought on by eating food that has been tainted. The Center for Disease Control and Prevention (CDC), USA, estimates that the *Escherichia coli* is a contributing factor in 76 million instances of foodborne disease [13]. The family Enterobacteriaceae includes *E. coli*. It is a short, Gram-negative, facultatively anaerobic, motile bacterium. One of the typical intestinal bacterial flora seen in poultry that is harmful to both people and animals is *E. coli* [7, 28]. Diarrheagenic *E. coli* are those that cause diarrhea and other enteric illnesses. Although many *E. coli* strains cause different intestinal illnesses, they colonize the intestines without causing any harm [40]. A kidney infection, septicemia, pneumonia, meningitis, hemorrhagic colitis, severe food poisoning, hemolytic uremic syndrome, abdominal pains, vomiting, dysentery, and bloody or non-bloody diarrhea are all possible effects of *E. coli* [22]. *E. coli* are naturally found in ruminants, such as sheep and goats. The infection also present on the animal's skin or in its feces at the time of slaughter, and it may transfer to the carcass by evisceration or skin removal process. Therefore, using subpar and unclean slaughter methods significantly raises the risk of *E. coli* contamination in meat [27]. The prevalence of multiple drug resistance of *E. coli* has increased due to the careless use of antibiotics. The use of antibiotics in the production of food animals has significant effects on human Health's. Additionally, *E. coli* is regarded as an indicator bacterium for Enterobacteriaceae family members' antibiotic resistance [30]. Many antibiotics may not be effective against *E. coli*. But *E. coli* isolates from poultry are typically antibiotic-resistant, especially they have widely utilized in the poultry business for a long time [12]. *E. coli* that is now a serious risk to both human and animal health is antibiotics-resistant to two and or more classes [21].

Materials and Methods

A total, 100 samples of meat or meat products were taken from the city of Udaipur. Two times every week, meat outlets and stores in the Rajasthani city of Udaipur were visited to gather samples of meat and RTE meat products. The samples were collected in a sterile container and brought to the lab in a refrigerated state using ice packs within two hours.

E. coli isolation and identification

E. coli were isolated from meat and or meat products using the procedure outlined by [29, 34]. For enrichment, the 25grams of sample was added to 225 ml of MacConkey broth after being properly triturated in a sterile mortar and pestle. A loopful of inoculum was plated on MacConkey agar and incubated at 37 °C for 24 hours after the culture flask had been cultured at 37 °C for 24 hours. The pink (lactose-fermenter) colonies were selected after 24 hours and streaked over eosin methylene-blue agar (EMB). Colonies with a metallic green gloss were chosen for additional verification.

Morphological characteristics

Gram negative bacilli that were grouped singly or in pairs were seen when the Gram-stained smear was examined under a microscope.

Biochemical evaluation

Numerous biochemical assays, including the indole test, methyl-red test, Voges-proskauer-test, citrate-utilization test, TSI test, sugar-fermentation test, catalase-test, oxidase-test, and urease-test, were carried out to confirm the suspected *E. coli* isolates [24, 31].

Antibiotic-susceptibility test

Antibiotic-susceptibility tests was performed on each and every *E. coli* isolate in accordance with [8]. The agar-disc diffusion method was used to test the susceptibility to an antibiotics. The test culture was isolated, and one isolated colony was injected in Luria-Bertani-broth and incubated for 24 hours at 37 °C. After that, a Mueller-Hinton agar plate was covered with the swab culture. The inoculated agar plate was covered with the antibiotic discs. To make sure that every disc made complete contact with the agar surface, each was

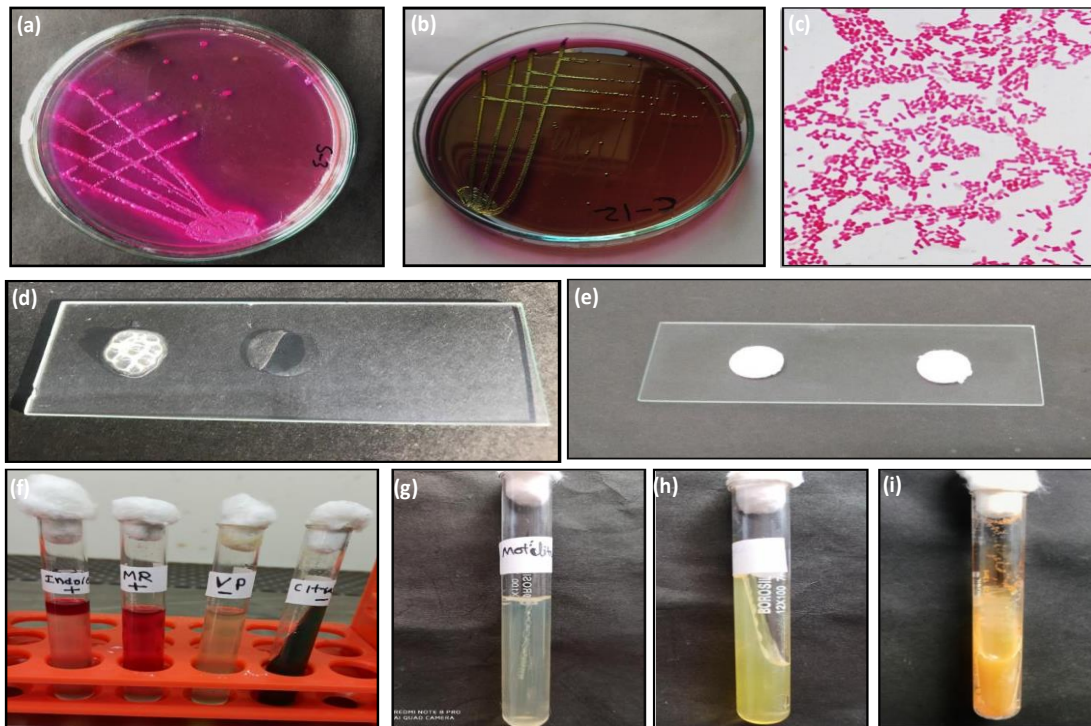
manually pressed down. A total of 10 antibiotic discs: amoxyclav; ampicillin; ceftriaxone; chloramphenicol; cotrimoxazole; enrofloxacin; erythromycin; gentamicin; nalidixic acid; and oxytetracycline. Following incubation, the diameter of the zone of inhibition was evaluated to ascertain the isolates' patterns of antibiotic susceptibility. According to the procedure outlined by [16, 26], the prevalence of multi-drug resistant *E. coli* was assessed.

Results

The test isolate that formed pink-colonies on MacConkey agar were chosen and streaked on EMB agar later. The 38 isolates with a green metallic sheen were identified from 100 samples and identified using Gram's staining and biochemical assays. The isolate was morphologically recognized as Gram negative- bacilli when the Gram's staining was done, and they were organized single or in pairs. Each of the 38 isolates underwent several biochemical assays after first isolation. In indole and methyl-red test, all suspicious isolates (n=38) developed a red ring on top and a red color, respectively. Similar to this, no growth of red or blue color was seen during the Voges-proskauer test or citrate test, respectively. This revealed that while citrate and the Voges-Proskauer test were negative for all of the suspicious isolates, they were all positive for the indole and MR tests. Fig. 1a-1i provide descriptions of the outcomes of several biochemical tests. All 100 samples of meat and RTE meat products, including chicken meat, mutton, chevon, and RTE meat products, were examined. Of these, 17, 9, 8, and 4 samples were found to be positive for *E. coli*, giving the prevalence rates of 56.65%, 45.00%, 40.00%, and 13.34%, respectively. *E. coli* was determined to be prevalent overall at 38.00% (38/100). The most effective antibiotics against the 38 isolates found in the various meat samples were chloramphenicol (84.22%), gentamicin (78.95%), ceftriaxone (78.95%), and ampicillin (63.16%). Erythromycin resistance was discovered in the isolates (94.73%). Additionally, moderately high resistance to oxytetracycline (36.45%) and amoxicillin/clavulanic acid (36.45%) was found in the current investigation. Table No. 2 displays the results of the antibiotic-susceptibility pattern of the *E. coli* isolates isolated from meat and RTE meat products.

Table 1: Antibiotic-susceptibility of *E. coli* isolates

Name of antibiotics	Chicken (17 isolates)			Mutton (9 isolates)			Chevon (8 isolates)			RTE products (4 isolates)		
	S	I	R	S	I	R	S	I	R	S	I	R
Gentamicin	13 (76.4%)	4 (23.5%)	0 (0%)	7 (77.8%)	1 (11.1%)	1 (11.1%)	6 (75%)	2 (25%)	0 (0%)	4 (100%)	0 (0%)	0 (0%)
Ceftriaxone	13 (76.4%)	2 (11.7%)	2 (11.7%)	7 (77.7%)	1 (11.1%)	1 (11.1%)	7 (87.5%)	0 (0%)	1 (12.5%)	3 (75%)	1 (25%)	0 (0%)
Nalidixicacid	44 (23.5%)	2 (11.7%)	11 (64.7%)	7 (77.7%)	1 (11.1%)	1 (11.1%)	6 (75%)	1 (12.5%)	1 (12.5%)	2 (50%)	2 (50%)	0 (0%)
Enrofloxacin	5 (29.4%)	6 (35.2%)	6 (35.2%)	8 (88.8%)	0 (0%)	1 (11.1%)	6 (75%)	1 (12.5%)	1 (12.5%)	4 (100%)	0 (0%)	0 (0%)
Ampicillin	8 (47.0%)	0 (0%)	9 (52.9%)	7 (77.7%)	0 (0%)	2 (22.2%)	6 (75%)	1 (12.5%)	1 (12.5%)	3 (75%)	1 (25%)	0 (0%)
Chloram-phenicol	11 (64.7%)	1 (5.8%)	5 (29.4%)	9 (100%)	0 (0%)	0 (0%)	8 (100%)	0 (0%)	0 (0%)	4 (100%)	0 (0%)	0 (0%)
Erythro-mycin	0 (0%)	0 (0%)	17 (100%)	0 (0%)	1 (11.1%)	8 (88.8%)	0 (0%)	1 (12.5%)	7 (87.5%)	0 (0%)	0 (0%)	4 (100%)
Amoxicillin/clavulanic Acid	5 (29.4%)	3 (17.6%)	9 (52.9%)	3 (33.3%)	3 (33.3%)	3 (33.3%)	6 (75%)	0 (0%)	2 (25%)	4 (100%)	0 (0%)	0 (0%)
Co-Trimoxazole	10 (58.8%)	2 (11.7%)	5 (29.4%)	8 (88.8%)	0 (0%)	1 (11.1%)	7 (87.5%)	0 (0%)	1 (12.5%)	3 (75%)	0 (0%)	1 (25%)
Oxy-tetracycline	6 (35.2%)	0 (0%)	11 (64.7%)	7 (77.7%)	0 (0%)	2 (22.2%)	7 (87.5%)	0 (0%)	1 (12.5%)	4 (100%)	0 (0%)	0 (0%)



Identification of *E. coli* : (a) Growth of the test culture on MacConkey agar plate. (b) Growth of the test culture on eosin methylene blue agar (EMB). (c) Gram's staining of the isolates (Gram negative rods). (d) Oxidase test (-ve). (e) Catalase test (+ve). (f) IMViC test of the isolates (++--). (g) Motility test of the isolates (h) TSI test (positive). (i) Urease test (negative) of the isolates.

According to our investigation, 56.66% of chicken meat samples included *E. coli*, which is comparable to the prevalence reported by [5, 11, 36] who reported 57.01%, 53.57%, 66.32%, and 61.76%, respectively. [17, 24, 38, 42] reported a significantly lower prevalence of 41.40%, 31.00%, 37.00%, and 34.00%, respectively. While a greater prevalence-rate of 93.75%, 83.5%, and 87.5%, respectively, was reported by [3, 18, 43]. Our study's somewhat high prevalence rate of *E. coli* in meat, and/or RTE meat products is a sign of unhygienic conditions surrounding the killing and sale of the animals. Additionally, unsanitary handling and shipping of meat products can be blamed for infection in those goods. Additionally, untrained and disorganized butchers produce and market the majority of the meat in the study area. This highlights the requirement to educate the public and butchers on hygienic meat production procedures.

Chloramphenicol (84.2%), Gentamicin (78.9%), Ceftriaxone (78.9%), and Ampicillin (63.1%) were the antibiotics most efficient against the 38 isolates found in the various meat samples. According to [2, 17, 33, 35], similar susceptibility values against chloramphenicol were 81.54%, 85.1%, 82.67%, and 67.1%, respectively. Conversely, [4, 19, 41] found that 79.4%, 73%, and 58% of the samples they tested showed resistance to chloramphenicol. Similar to this, [17, 33] found that antibiotic-sensitivity to gentamicin was 85.7% and 81.0%, respectively. The resistance to gentamicin was found to be 46.6% and 57.47%, respectively, by [1, 20]. Ceftriaxone was shown to be effective to *E. coli* isolates in 73% and 94.29% of the cases, respectively, according to [23, 33]. The effectiveness of ampicillin against 52.9% of the isolates was also reported by [2]. Although [10, 16, 19, 25, 37] found that resistance to ampicillin was seen to be 57%, 84.93%, 98%, 80.43%, 63.4%, 62%, and 75.6%, respectively. The erythromycin resistance of the *E. coli* isolates isolated. However, there was also evidence of fairly high resistance to oxytetracycline and amoxicillin + clavulanic acid. The widespread and careless use of

antibiotics to treat illnesses in humans and promote animal growth is highlighted by this rising resistance. A significant source of ESBL-producing *E. coli* for humans is poultry and chicken meat. These drug-resistant *E. coli* can directly or indirectly infect people through food, and they may also spread resistance genes to susceptible bacteria. In order to reduce the risk to the public health posed by the growth of antibiotic-resistant bacteria, careful use of antibiotics is important.

Conclusion

In this investigation, 100 samples of meat and RTE meat products were gathered from various retail meat stores in Udaipur, and the prevalence in *E. coli* was reported to be 38% (38/100). Total *E. coli* isolates found in chicken meat, mutton, chevon, and RTE meat products were 17, 9, 8, and 4, respectively, with prevalence rates of 56.66%, 45%, 40%, and 13.33%. Out The most effective antibiotics for the 38 isolates recovered from the various meat samples were ampicillin (63.15%), gentamicin (78.94%), chloramphenicol (84.21%), and ceftriaxone (78.94%). Erythromycin resistance was discovered in the isolates (94.73%). Additionally, in the current investigation, moderately high resistance to oxytetracycline (36.84%) and amoxicillin/clavulanic acid (36.84%) was observed. These findings highlight the necessity of using antibiotics sparingly in animal husbandry and the threat to the general public's health posed by the proliferation of bacteria that are resistant to them and cause food-borne illnesses.

References

1. Abd ET, Ammar A, AM, Nasef SA, Reda RM. Antibacterial resistance and resistance gene detriments of *E. coli* isolated from chicken. Benha Veterinary Medical Journal. 2015;28(2):231-240.
2. Adelaide OA, Bii C, Okemo P. Antibiotic resistance and

- virulence factors in *E. coli* from broiler chicken slaughtered at Tigoni processing plant in Limuru, Kenya. *East Afr. Med J.* 2008;85(12):597-606.
3. Albarri OM, VarI, Meral M, Heshmati B, Koksai F. Prevalence of *Escherichia coli* isolated from meat, chicken and vegetable samples in Turkey. *Journal of Biotechnology Science Research.* 2017;4(3):214-222.
 4. Amer MM, Mekky, HM, Amer AM, Fedawy, HS. Antimicrobial resistance genes in pathogenic *Escherichia coli* isolated from diseased broiler chickens in Egypt and their relationship with the phenotypic resistance characteristics. *Veterinary World.* 2018;11(8):1082-1088.
 5. Ashraf N, Hussain I, Siddique F. Microbial burden and drug residual analysis in raw meat samples from different towns of Faisalabad, Pakistan. *Scholar's Adv. Anim. Vet. Res.* 2015;2(4):228-237.
 6. Basak S, Singh P, Rajurkar M. Multidrug resistant and extensively drug resistant bacteria: A study. *Journal of pathogens;* c2016.
 7. Bashar T, Rahman M, Rabbi FA, Noor R, Rahman MM. Enterotoxin profiling and antibiogram of *Escherichia coli* isolated from poultry feces in Dhaka District of Bangladesh. *Stam ford Journal of Microbiology.* 2011;1(1):51-57.
 8. Bauer AW, Kirby WM, Sherris JC, Truck M. Antibiotic susceptibility testing by standardized single disk method. *American Journal of Clinical Pathology.* 1966;45(4):493-496.
 9. Beyi AF, Fite AT, Tora E, Tafese A, Genu T, Kaba T, *et al.* Prevalence and antimicrobial susceptibility of *Escherichia coli* O157 in beef at butcher shops and restaurants in central Ethiopia. *BMC Microbiology.* 2017;17(1):49.
 10. Bhave S, Kolhe R, Mahadevaswamy R, Bhong C, Jadhav S, Nalb andS *et al.* Phylogroupin and antimicrobial resistance analysis of extra intestinal pathogenic *Escherichia coli* isolated from poultry species. *Turkish Journal of Veterinary and Animal Sciences* 2019;43(1):117-126.
 11. Bhoomika SS, Patyal A, Gade NE. Occurrence and characteristics of extended-spectrum beta-lactamases producing *E. coli* in foods of animal origin and human clinical samples in Chhattisgarh. *India Veterinary World.* 2016;9(9):996-1000.
 12. Blanco JE, Blanco M, Mora A, Croas C, Blanco J. *Escherichia coli* associated with coli Septicaemia in Spain. *Medicina Veterinaria.* 1996;13(12):680-686.
 13. Centers for Disease Control and Prevention (CDC). Updateonmulti-state outbreak of *E. coli* O157: H7 infections from fresh spinach; c2006.
 14. Chavhan SK, Dewanand RK, Anshuja AN. Pathogenic attribute of *Escherichia coli* isolated from commercial broilers. *Indian Vet. J.* 2012;89(1):39-40.
 15. Davis GS, Waits K, Nordstrom L, Grande H, Weaver B, Papp K, *et al.* Antibiotic resistant *Escherichia coli* from retail poultry meat with different antibiotic use claims. *BMC Microbiology.* 2018;18(1):174.
 16. Dsani END. Antimicrobial resistance patterns of *Escherichia coli* isolated from beef, mutton and chevon in the greater Accra region of Ghana (Doctoral dissertation, university of GHANA); c2019.
 17. Equar Y. Characterization of Drug Resistance Patterns of *E. coli* isolated from milk collected from small scale dairy farms reared in Holeta and Burayu, and meat from Addis Ababa abattoirs enterprise and Alema farms laughter slab (Doctoral dissertation, Addis Ababa university); c2016.
 18. Eyy A, Arslan S. Prevalence of *Escherichia coli* in retail poultry meat, ground beef and beef. *Med. Weter,* 2012.68(4):237-240.
 19. Farhan MB, Jubeer OJ. Molecular study of some virulence factor Sentero-pathogenic *Escherichia coli* isolated from new-born till the age of one year. *Journal of Pharmaceutical Sciences and Research.* 2018;10(12):3388-3392.
 20. Gai W, Wang J, Wang J, Cui Z, Qu Z, Cui J, *et al.* Molecular classification and drug resistance analysis of *Escherichia coli* isolated from poultry in China. *International Journal of Clinical and Experimental Medicine.* 2015;8(1):836-844.
 21. Gonzalez EA, Blanco J. Serotypes and antibiotic resistance of Vero-toxigenic (VTEC) and necrotizing (NTEC) *Escherichia coli* strains isolated from calves with diarrhoea. *FEMS Microbiology Letters.* 1989;60(1):31-36. 22.
 22. Gupta B, Ghatak S, Gill JPS. Incidence and virulence properties of *E. coli* isolated from fresh fish and ready-toeat fish products. *Veterinary World.* 2013;6(1):5-9.
 23. Gupta MD, Sen A, Das A. Occurrence of *Escherichia coli* carrying shiga toxin producing genes in buffaloes on small holdings in Bangladesh. *Veterinary World.* 2018;11(10):1454-1458.
 24. Ibrahim WA, Marouf SA, Erfan AM, Nasef SA, El Jakee JK. The occurrence of disinfectant and antibiotic-resistant genes in *Escherichia coli* isolated from chickens in Egypt. *Veterinary World.* 2019;12(1):141-145.
 25. Kaushik P, Anjay, Kumari S, Dayal S, Kumar S. Antimicrobial resistance and molecular characterization of *E. coli* from poultry in Eastern India. *Veterinarian Italiana.* 2018;54(3):197-204.
 26. Magiorakos AP, Srinivasan A, Carey RB, Carmeli Y, Falagas ME, Giske C *Getal.* Multi-drug resistant, extensively drug-resistant and Pandrug-resistant bacteria: an international expert proposal for interim standard definitions for acquired resistance. *Clinical Microbiology and Infection.* 2012;18(3):268-281.
 27. McEvoy JM, Doherty AM, Sheridan JJ, Thomson-Carter FM, Garvey P, McGuire Letal. The prevalence and spread of *Escherichia coli* o157: H7 at a commercial beef abattoir. *Journal of Applied Microbiology.* 2003;95(2):256-266.
 28. Melnick JL, Adelberg EA. Review of medical microbiology, California. 16th Edn. Lange Medical Publication; c1987. p. 122-144.
 29. Quinn PJ, Markey BK, Leonard FC, Hartigan P, FanningS, Fitzpatrick E. *Veterinary Microbiology and Microbial Disease;* c2011.
 30. Rahman MA, Rahman AKMA, Islam MA, Alam MM. Antimicrobial resistance of *Escherichia coli* isolated from milk, beef and chicken meat in Bangladesh. *Bangladesh Journal of Veterinary Medicine.* 2017;15(2):141-146.
 31. Rajput SK, Gururaj K, Tiwari U, Singh G. Study of the characterization of *E. coli* isolates in goat kids. *Indian Res. J Genet. & Biotech.* 2014;6(1):324-329.
 32. Rodriguez-Cavallini E, Rodriguez C, Gamboa MM, Arias ML. Microbiological evaluation of ready to eat foods manufactured by small Costa Rican industries. *Archivos Latino americanos De Nutricion.* 2010;60(2):179-183.
 33. Sahoo TK, Sahoo L, Sarangi LN, Panda SK, Panda HK.

- Prevalence, isolation, characterization and antibiogram study of pathogenic *Escherichia coli* from different poultry farms of Odisha. *Journal of Advanced Veterinary Research*. 2012;2(3):169-172.
34. Sarba EJ, Kelbesa KA, Bayu MD, Gebremedhin EZ, Borena BM, Teshale A. Identification and antimicrobial susceptibility profile of *Escherichia coli* isolated from backyard chicken in and around ambo, Central Ethiopia. *BMC Veterinary Research*. 2019;15(1):85.
 35. Sharada R, Ruban SW, Thiyaeeswaran M. Antibiotic resistance pattern of *Escherichia coli* isolated from poultry in Bangalore. *International Journal of Microbiology*. 2009;7(1):1-5.
 36. Sharma M, Singh DP. Recovery of bacterial contaminants from various foods in the state of Himachal Pradesh, India FAVA- OIE joint symposium on emerging diseases; c2008.
 37. Subedi M, Luitel H, Devkota B, Bhattarai RK, Phuyal S, Panthi P, *et al.* Antibiotic resistance pattern and virulence genes content in avian pathogenic *Escherichia coli* (APEC) from broiler chickens in Chitwan, Nepal. *BMC Veterinary Research*. 2018;14(1):113.
 38. Vazgecer B, Ulu H, Oztan A. Microbiological and chemical qualities of chicken doner kebab retailed on the Turkish restaurants. *Food Control*. 2004;15(4):261-264.
 39. World Health Organization (WHO). The role of food safety in health development, report of joint FAO/WHO Expert Committee on Food Safety, Geneva; c1984.
 40. Xia X, Meng J, McDermott PF, Ayers S, Blickenstaff K, Tran TT, *et al.* Presence and characterization of Shiga toxin-producing *Escherichia coli* and other potentially Diarrheagenic *E. coli* strains in retail meats. *Appl. Environ. Microbiol.* 2010;76(6):1709-1717.
 41. Xie M, Lin D, Chen K, Chan EWC, Yao W, Chen S. Molecular characterization of *Escherichia coli* strains isolated from retail meat that harbor bla CTX-M and fosA3genes. *Antimicrobial Agents and Chemotherapy*. 2016;60(4):2450-2455.
 42. Zargar HK. Evaluation of hygienic quality of raw meat (Mutton and chicken) and characterization of isolated pathogens (Doctoral dissertation, Division of Veterinary Public Health and Epidemiology, Sher-e-Kashmir University of Agricultural Sciences and Technology of Jammu); c2016.
 43. Zhao S, Blickenstaff K, Bodeis-Jones S, Gaines SA, Tong E, McDermott PF. Comparison of the prevalence and antimicrobial resistances of *Escherichia coli* isolates from different retail meats in the United States, 2002 to 2008. *Appl. Environ. Microbiology*. 2012;78(6):1701-1707.