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A comparative study of the immune responsiveness in native chickens of Chhattisgarh, PB2 and their crosses under intensive system

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Abstract

Immunity is a very important trait in rural poultry. The present experiment was a comparative study of the immune responses of local native chickens (T₁), PB2, a colour broiler (T₂), and crossbreed F1 (T3). In the study of cell-mediated immunity that is evoked by PHA-P injection, immunity values in terms of foot index for T₁, T₂, and T₃ were found to be $0.08\pm.03$, $0.10\pm.02$ and $0.11\pm.01$, respectively. In the study of humoral immunity, a 1% sheep RBC solution was injected. After 5 days, the mean HA titres of T₁, T₂ and T₃ were found to be 4.5 ± 0.2 , 8.33 ± 1.0 and 8.16 ± 0.4 respectively. A T₁ 4th day mean titers of T₁, T₂ and T₃ were found to be 7.0 ± 0.2 , 10.16 ± 0.4 , and 10.0 ± 0.2 respectively. On the 5th and 14th day significantly lower immune response was reported in T₁ whereas T₂ and T₃ were not differed statistically. At 21th day, mean HA titres of T₁, T₂ and T₃ were found as 5.16 ± 0.6 , 5.50 ± 1.0 and 6.33 ± 1.2 respectively.

Keywords: Cell mediate immune response, humoral immunity, native chicken, PB2, crossbreed, PHAP, SRBC

1. Introduction

The Indian government and research institutes are continuously developing and propagating improved varieties of chicken for sustainable production in intensive and backyard systems. New poultry varieties are exhibiting higher productivity but are facing the disease outbreak, even though vaccination programmes are in place. Hence, improvements in immunity have enduring effects on the population. Disease resistance in poultry is controlled by several genes. Disease resistance is important for maintaining health and protecting the chicken from various pathogenic organisms. Disease resistance traits of different breeds and varieties have been studied; they are very important in breed development programs. Disease resistance can be analysed by studying the immune status of chickens. It is called immunocompetence traits. The sheep RBCs act as a foreign antigen for chickens. It is used for assessing the humoral response of an individual without affecting their health. After inoculation of sheep RBC, it produces specific antibody titres (anti-SRBC), which are used as a tool to determine the humoral immune response. It was found that individuals with a higher SRBC response also reported higher antibodies against Marek's disease, Ranikhet disease virus, and coccidia. Humoral immune response is moderately heritable, so it is important for disease resistance, increased vaccine efficacy, and the health status of farms. And in vivo mitogen PHA-P to quantify the lymph proliferative cell-mediated immune response.

2. Material and methods

The present investigation was carried out at the College of Veterinary Science and A.H. Anjora, Durg Chhattisgarh, on local native chickens (T_1) , PB₂ colour broilers (T_2) , and F1 (crosses of native male and PB₂ female) birds (T_3) . 240 chicks were reared in each group for 14 weeks. At the 10th week for cell-mediated immune response evaluation, 6 birds per group were selected randomly that were free from disease, and similarly, 6 birds per group were selected for the humoral immune response study.

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2.1 Humoral Immune Response

The immune response of the birds was determined by haemagglutination titres against sheep red blood cells (SRBC). SRBC acts as an antigen. (Rajkumr *et al.*, 2011) ^[1]. 20 ml of blood from a sheep was taken in an equal amount of Alsever's solution. Blood collected in Alsever's solution had been kept in a plastic test tube after removing the supernatant. Alsever centrifuged it at 2500 rpm for 10 minutes, then washed it three times in phosphate-buffered saline solution till the supernatant was cleared. 1 ml of sheep RBC (PCV) was mixed in 99 ml of PBS solution to make a 1% sheep RBC suspension and kept in the refrigerator at 4 ^oC °C until further usage.

6 birds per group were injected with one ml of 1% sheep RBC suspension intravenously at 10^{th} weeks of age. One ml of blood was collected on 5^{th} , 14^{th} and 21^{st} day of post-immunization. The serum was then separated out from the blood and kept at -20 °C until further use. The heamaglutination test for sheep RBC was performed as per procedure. After the HA test, the reciprocal of the highest dilution, which appears to be clear agglutination, was the end titer. Titers wrote as log 2.

2.2 Cell mediated immune response

The *in vivo* cell-mediated immune response to phytohaemagglutinin type P was determined by the method of Cheng and Lamont (1998) ^[2]. Phytohacmagglutinin type P elicits immune responses influenced by a subpopulation of 1-helper and T-suppressor cells. An individual that responds better to PHA-P has a higher level of cellular immunity,

influencing T-cell mechanisms that prevent lymphoma formation. Six birds per group were taken to evaluate the response of PHA-P. The procedure was as follows:

PHA-P (0.1 mg/ 0.1 ml PBS) was inter-digitally injected between the 3^{rd} and 4^{th} toes of the right shank of the bird. The left leg was chosen as the control and was injected with 0.1 ml PBS. The skin index (foot web index) was calculated as the difference between the swellings (measured by a micrometre instrument) in the right and left legs before and 24 hours after injection.

Foot index FI (mm) = (Post inj.-Pre inj.) - (Post PBS-Pre PBS)

2.3 Statistical analysis

To see the difference between different treatment groups, a one-way analysis of variance was applied (Snedecor and Cochron 1994)^[3]. If there is any significant difference in any group, then DMRT is applied (Steel and Torrie 1984)^[4].

3. Result

3.1 Cell mediated immunity

The mean cell-mediated immune responses of $T_1 T_2$ and T_3 are presented in table 4.36 and depicted in Figure 4.36 Non significant effect of immunity was reported among the groups; the immunity value in terms of foot index for T_1 , T_2 , and T_3 were found to be $0.08\pm.03$, $0.10\pm.02$ and $0.11\pm.01$ respectively.

Age	Local Native	PB-2	Native male X PB2 female (at Farm)	P Value
	$0.08 \pm .03$	$0.10 \pm .02$	0.11±.01	NS

3.2 Humoral immunity

Post inoculation	Local Native	PB-2	Native male X PB2 female (at Farm)	P Value
5 th day	$4.5 \pm .28^{a}$	8.33±1.01 ^b	$8.16 \pm .44^{b}$	*
14 th day	7.0±0.28 ^a	10.16±0.44 ^b	10.0±0.28 ^b	**
21 day	5.16±0.66	5.50±1.04	6.33±1.20	NS

The mean humoral immune responses of T_1 , T_2 and T_3 are presented in table 4.37. In the study of humoral immunity, a 1% sheep RBC solution was inoculated. Immune responses were measured at 5th, 14th and 21th days post-inoculation by the heamaglutination test. In the experiment at 5th day a significant lower immune response was reported in T₁, whereas no statistical difference was found between T₂ and T₃, at 5th day mean titre of T_1 , T_2 and T_3 were found as 4.5±.28, 8.33±1.01 and 8.16±.44 respectively. At 14^h days, the mean titre of T₁, T₂ and T₃ were found as $7.0\pm0.28m$, 10.16 ± 0.44 and 10.0±0.28 respectively. At 5th and 14th day significant lower immune was found in T_1 . Whereas T_2 and T_3 were not differed from each other. On 21st day, no significant difference in titre was reported among groups. Mean titre of $T_1,\ T_2$ and T_3 were found as $5.16\pm0.66,\ 5.50\pm1.04$ and 6.33±1.20 respectively. The decreasing titre values were found in the 21st day.

4. Discussion

Similar to present result Singh and Singh (2004)^[5], Chatterjee *et al.* (2007)^[6], Divya *et al.* (2018)^[7], and Sharma *et al.* (2020)^[8] were reported no significant effect on cell mediate immunity among different breeds. Pathak *et al.* (2017)^[9] measured higher foot web thickness in aseel. In present study

local native's immune response after SRBC inoculation was reported to be lower as compared to Kadaknath and Aseel (Kundu et al., 1999), Chatterjee et al., 2007) ^[10, 6], contrary to this lower value reported in Kadaknath birds (Saxena et al., 2012) ^[11]. whereas in the present study the titre of native was similar to findings Radhika et al. (2017) ^[12], Higher values were found in F1 and PB2 as compared to Aseel and Kadaknath (Kundu et al., 1999) ^[10], in the present study of PB2 the result of immune response is in accordnce with Prasad *et al.* (2008) ^[13] findings in Gramapriya.In the present study immune response value of F₁ crossbreed is in close agreement with Divya et al. (2018) [7] findings in Aseel crosses. A similar result was also obtained by Barik et al. (2018) ^[14] in Vanraja. And Alam et al. (2021) ^[15] in NNRIR (progeny of naked neck crossed with RIR) In the present study, local native chickens have less immunity as compared to indigenous breeds Assel and Kadaknath. The reason might be that these breeds are known for their hardiness and disease resistance; they have developed natural immunity over the generations, which may be stronger than local breeds. In intensive systems, chickens are kept in close proximity and natural behaviours of local chickens, like dust bathing, preening, and foraging, which increase the risk of disease transmission. Local chickens may not have developed

immunity to the specific pathogens present in intensive settings; constant exposure to pathogens and intensive environmental stress make their immune systems weak. In the study of cross-breed chickens, higher immunity was reported. The reasons might be hybrid vigour, genetic diversity and complementary gene combinations. The colour broiler shows more immunity because it may have greater genetic diversity compared to the commonly used white-feathered variety. Selective breeding is also a reason for better immunity.

5. Conclusion

In the present study under intensive system humoral immune response was found better in F_1 crossbred and PB_2 whereas lower imuunity response obtained in local native because native chicken of Chhattisgarh is less immune against pathogen found in intensive system, whereas antibody titre was reported declining trend after 14th day post injection of sheep RBC.

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