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Oxidative stress and its effects on reproductive performance in thermally-stressed ewes

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Abstract

This paper reviewed the impact of thermally-induced oxidative stress on reproduction in ewes. Thermally-induced oxidative stress is associated with excess generation of reactive oxygen species (ROS) during aerobic respiration and metabolic activities within the body. Excess ROS have been implicated in pathophysiologic mechanisms impairing reproductive processes, including changes in cyclic luteal and endometrial processes, poor follicular development, ovulation, fertilisation, embryogenesis, embryonic implantation, and placental differentiation and foetal growth. High levels of ROS during pregnancy result in pregnancy disorders such as spontaneous abortions, embryopathies, pre-eclampsia, foetal growth restriction, pre-term abortion and low-birth weight. Thermally-induced stress occurs as a result of extreme cold and hot temperatures, and its impact on animals is determined by nature of management practices being used in the farm. Heat stress suppresses reproductive hormonal expression and activities, resulting in poor reproductive performances, including the exhibition of poor oestrus behaviour and mating ability; while cold stress suppresses reproductive neuroendocrine axis, which adversely affects conception rates and mating behaviours. Other oxidative stress factors exerting negative effects on reproduction are poor nutrition, solar radiation, environmental pollutants, high ambient temperature and high relative humidity, expressed as temperature-humidity-index. The stressors contribute to adverse pregnancy outcomes, increase in the susceptibility of offspring to disease by compromising the offspring immune status and overall reproductive performance in ewes. The link between oxidative stress and reproductive processes such as spermatogenesis, oogenesis, fertilisation, embryonic development, implantation and foetal growth are critical therapeutic and prophylactic interventions in reproductive medicine. The role of oxidative stress in reproductive processes and the state of knowledge on the association between oxidative stress and pregnancy outcomes in ewes are elucidated. It is concluded that oxidative stress is an important silent condition undermining reproductive performance in sheep resulting in serious economic loss. Reduction of its impacts will enhance productivity and performance in ewes.

Keywords: Environmental stressors, oxidative stress, reactive oxygen species, reproduction, pregnancy, oestrus cycle, ewes.

Introduction

Sheep production form an important component of the livestock economy among rural populations in Africa, including Nigeria. Apart from their socio-cultural roles as sources of meat and food security, they are important rural financial resources, serving as cash savings and are used in festive activities. Sheep contribute 11% of the meat industry in Nigeria. Their production is an important component of livestock production in many parts of the world, where they are raised for mutton, milk, wool and hides and skins. In Australia, sheep production is an important source of mutton, wool and milk. In the Mediterranean countries, sheep milk is rated higher for nutritive value and cheese processing outputs than cow milk. Therefore, sheep milk is more expensive than cow milk. The Mediterranean countries have a long history of consumption of sheep milk as they associate it with longevity. Thus, sheep production forms an important part of the integrated agricultural economy across the world ^[1]. Various management systems are employed in sheep production in tropical environments. In Nigeria, traditional management system such as nomadism, transhumance and agropastoralism are among the common methods used, where animals are raised and fed through herding and open grazing and feeding of animals on crop-residue with little or no supplemental feeding ^[2].

The production systems expose animals to harsh, tropical climatic conditions, presenting seasonal variability in feed quality and supply often resulting in nutritional stress. The experience extreme weather conditions throughout the year, predisposing them environmental stressful conditions and diseases. Nutritional stress, environmental stress and diseases compromise the productivity of livestock and, especially, their reproductive performance. Nutritional stress is associated with underfeeding due to the seasonal variation in feed quality and availability that brings about roaming and scavenging of food through nomadism and transhumance practices. Consequently, there is an increase in the incidence of reproductive disorders and infertility in animals [3, 4, 5] and in women [6].

Environmental conditions precipitate thermal (hot and cold) stress as the result of extreme weather conditions due to climatic changes, which subject the animals to oxidative stress that undermine the reproductive performance. Oxidative stress is also associated with tropical diseases especially ecto- and endo-parasitism which predispose animals to changes in the vulgaris of weather such as changes in ambient temperature and humidity. In tropical countries this situation breaks the weather into two main weather conditions, viz: hot and dry seasons. Both seasons are characterized by variations in disease burdens on animals due to differences in changes in management systems, accessibility of good nutrition and water, thus, predisposing them to oxidative stress. The un-favorable conditions associated with management systems such as nomadism and transhumance practices cause varying levels of oxidative stress, which undermine the productive capacity of ewes and result in poor reproductive performance as well as huge economic loss. Measures aimed at ameliorating oxidative stress in sheep production are, thus, crucial for the sustainability of their production in tropical environments. Such measures are required to interfere in the pathophysiologic mechanisms underlying oxidative stress resulting in excess generation of ROS, which adversely affect reproductive performance in ewes. This paper reviews oxidative stress associated with changes in environmental conditions and its adverse effects on reproductive performance in sheep exposed to thermal stress.

Oxidative Stress

Oxidative stress (OS) plays a crucial role in the mechanism of diseases arising from extreme environmental conditions. It occurs as result of imbalances in the redox (reduction and oxidation) reactions due to the production and scavenging of ROS [7, 8]. Redox reactions involve the transfer of electrons between molecules during metabolic and aerobic respiration due to cellular response to endogenous and exogenous stimuli [7, 8, 9]. Excessive endogenous stimuli such as disease conditions, increased aerobic respiration and metabolic activities; and exogenous stimuli including environmental disrupting substances, thermal stress, wind speed and solar radiation. These stimuli increase the generation of reactive species, comprising oxygen and nitrogen free radical species as by-products, resulting in oxidative stress [7, 8, 10]. Although reactive species play a central role in normal cellular function in providing an important feed-back loop between metabolic activity and regulation of cellular functions, excess production of ROS in the body exerts deleterious effects in the body and overall performance [11]. The ROS arise due to an imbalance between pro-oxidants and antioxidants in stressful conditions, as a result of increased antioxidant utilisation, impaired

buffering capacity or immune dysfunction [8, 11]. Though several reactive molecules are produced in the body, ROS and reactive nitrogen species (RNS) are the most important. ROS comprise superoxide, hydroxyl radical, peroxy radical, while the RNS include nitrogen oxide and peroxy nitrite. These species are generated continuously in small quantities during normal aerobic respiration and metabolic activities in the body. The reactive species are involved in several physiological processes, including protein phosphorylation, activation of transcription factors, cell differentiation, apoptosis, oocyte maturation, steroidogenesis, cell immunity and cellular Defence against pathogens [11, 12]. The effects of ROS on cells become deleterious when they are generated in excess, resulting in oxidative stress, when there is an imbalance between ROS and antioxidant activities. Oxidative stress is common in animals exposed to extreme environmental conditions such as excessive thermal, wind speed and solar radiation [8].

Heat stress impairs animal performance, and has been implicated in promoting oxidative stress, either through excessive ROS production or decreased antioxidant defenses [13]. Some diseases like sepsis, ketosis, mastitis, and enteritis, and pneumonia, respiratory and joint diseases are associated with excess ROS [11]. Excess ROS generation also causes infertility and poor reproductive performance in animals [14]. The significant roles of ROS in the pathogenesis of reproductive disorders have been reported and these include attack on the fluidity of the sperm plasma membrane and DNA integrity in the sperm nucleus; induced DNA damage which may accelerate the process of germ cell apoptosis, leading to the decline in sperm counts associated with male infertility [15, 16, 17].

In female, ROS mediated the pathophysiology of endometriosis and unexplained infertility with high follicular fluid ROS levels, resulting in negative IVF outcomes, apparently responsible in *hydrosalpinges* fluid mediated embryo toxicity as well as poor *in vitro* embryonic development. In the female, oxidative stress has been reported to influence oocyte development, maturation, follicular atresia, *corpus luteum* function and luteolysis [18]. Though ROS may play a significant role in the modulation of gamete interaction and successful fertilization, excess ROS reduce oocyte and embryo quality both *in vitro* and *in vivo*. The ROS exert considerable impact on post-fertilisation development, such as cleavage rate, blastocyst yield and quality, which are indicators of assisted reproduction outcomes [19]. Oxidative stress has been reported to cause embryo resorption, placental degeneration with subsequent alteration in maternal-foetal exchanges, delay in foetal growth, pregnancy interruption, and still-births during pregnancy [20]. Excess ROS generation modulate the microenvironment in the female, resulting in follicular, *hydrosalpinges* and peritoneal fluid accumulation. It interferes with implantation and early embryo development, and causes embryo toxicity due to poor *in vitro* embryonic development, resulting in poor pregnancy outcomes [15, 21]. Oxidative stress is implicated in pathophysiology of endometriosis and unexplained infertility, and high concentrations of ROS in follicular fluid is reported to be associated with negative *in vitro* fertilisation (IVF) outcomes [15]. The ROS also cause DNA damage, resulting in various genomic defects due to telomere shortening; for example, aplastic anaemia in the bone marrow and progressive fibrosis in the lungs and liver [22, 23, 24]. The ROS undermine the development of germ cells, which adversely affects the quality of ovum production. They also induce oocyte

implantation disorders, endometriosis, embryo resorption, placental degeneration with subsequent alteration in maternal-foetal exchanges, delay in foetal growth, pregnancy interruption and stillbirths in sheep [20, 25, 26]. In female animals, oxidative stress has been associated with irregular oestrous cycle, polycystic ovary syndrome, endometritis, infertility, poor embryo development and pregnancy failure [14]. Pregnancy, especially at the last trimester and early lactation has been associated with oxidative stress due to higher nutrient demands for rapid foetal growth and the production of large amounts of colostrum and milk [27]. The ROS exert negative impact on weight gain and nutrient utilisation in ewes, especially during the last trimester of pregnancy, and at the onset of lactation, when the body requirements for micronutrients and antioxidants are very high, but the supply is grossly inadequate [28].

The indices of oxidative stress have a strong correlation with infertility in the females. The ROS levels and selenium-dependent glutathione peroxidase activity in follicular fluid are directly correlated with pregnancy rates, while lipid peroxidation, total antioxidant capacity, and superoxide dismutase activity have strong correlation with oocyte fertilisation and pregnancy rates, following IVF and other assisted-reproductive techniques [21]. However, the oxidative stress maker in granulosa cells, 8-hydroxy-2-deoxyguanosine shows negative correlation with embryo quality following IVF [21].

Oxidative stress is associated with infertility in rams and other male animals. Normal ROS levels modulate the fertilizing capabilities of the spermatozoa, but excess ROS above the physiological levels result in structural and functional damage, which may lead to motility loss, premature acrosomal reaction, lipid peroxidation, apoptosis, and DNA damage [18, 21]. Oxidative by-products generated by ROS include 8-oxo-7,8-dihydroxyguanosine, which cause lipid peroxidation, RNA transcripts damage and cause DNA fragmentation with a resultant mutagenic effect and overall negative effects on sperm quality [29]. The compound also causes negative impact on the success rates of assisted-reproductive techniques such as artificial insemination (AI), *in vitro* fertilisation (IVF) and intracytoplasmic sperm injection (ICSI) [30]. Oxidative stress reduces sperm count due to germ cell apoptosis, resulting in male infertility in animal models and humans [15, 31, 32, 33]. Furthermore, it has been reported that the generation of ROS, including superoxide anion, hydrogen peroxide and hydroxyl radicals during oxidative stress, increases the decay of sperm membrane lipid and DNA, and increases DNA strand breaks [30]. Damages to

sperm membrane and DNA by excess ROS result in necrozoospermia, asthenozoospermia, and DNA fragmentation in sheep, man and other mammals [21, 32, 33, 34, 35]. Excess ROS impair overall sperm production and function due to oxidative damage to somatic cells and spermatozoa [21, 23, 31, 36]. They are associated with erectile dysfunction, sperm damage and motility in animals and men [37, 38].

Reactive Oxygen Species and Reproductive Performance in Sheep

Thermal stress comprising heat and cold stress is associated with oxidative stress and poor reproductive performance in ruminants [39, 40].

Heat and cold stress are very serious conditions that occur when the body can no longer maintain a normal temperature. They result in serious illnesses and injuries, permanent tissue damage or death. Heat stress is associated with tropical or hot environments, while cold stress is associated with cold or temperate environments. In tropical climates such as occur in Nigeria, seasonal climatic variations exist, often with extreme weather variations. The two main seasons that could be classified in most northern parts of Nigeria are the dry (November – May) and rainy seasons (June – October). The dry season has no rain, and, therefore, lacks precipitation. It is further classified into cold-dry with hazy (harmattan) winds (November – February), and extreme hot period (March – May) [41]. The rainy season has rainfalls and, therefore, higher precipitation and there may be high relative humidity and high temperature (June – August) or high relative humidity and low ambient temperatures (September - October) [41]. The resultant temperature-humidity index (THI), which is an expression of heat load, has deleterious effects on overall sheep performance [42].

Cold stress is induced by low temperatures, high wind speed, high relative humidity, and/or contact with cold water or surfaces, while heat stress is characterized by high temperatures, low wind speed and moderate/high relative humidity. A cold environment forces the body to function harder to maintain its temperature as it draws heat from the body, while a hot environment results in the body functioning harder to dissipate heat. Oxidative stress adversely affects sheep productivity especially in both temperate and tropical environments. The high prevalence of pests and diseases, solar radiation, thermal and nutritional stress are important predisposing factors to oxidative stress in ruminants. Multiple stresses acting concomitantly are reported to exert more severe negative effects on all reproductive parameters than individual stress factors (Fig. 1) [43].

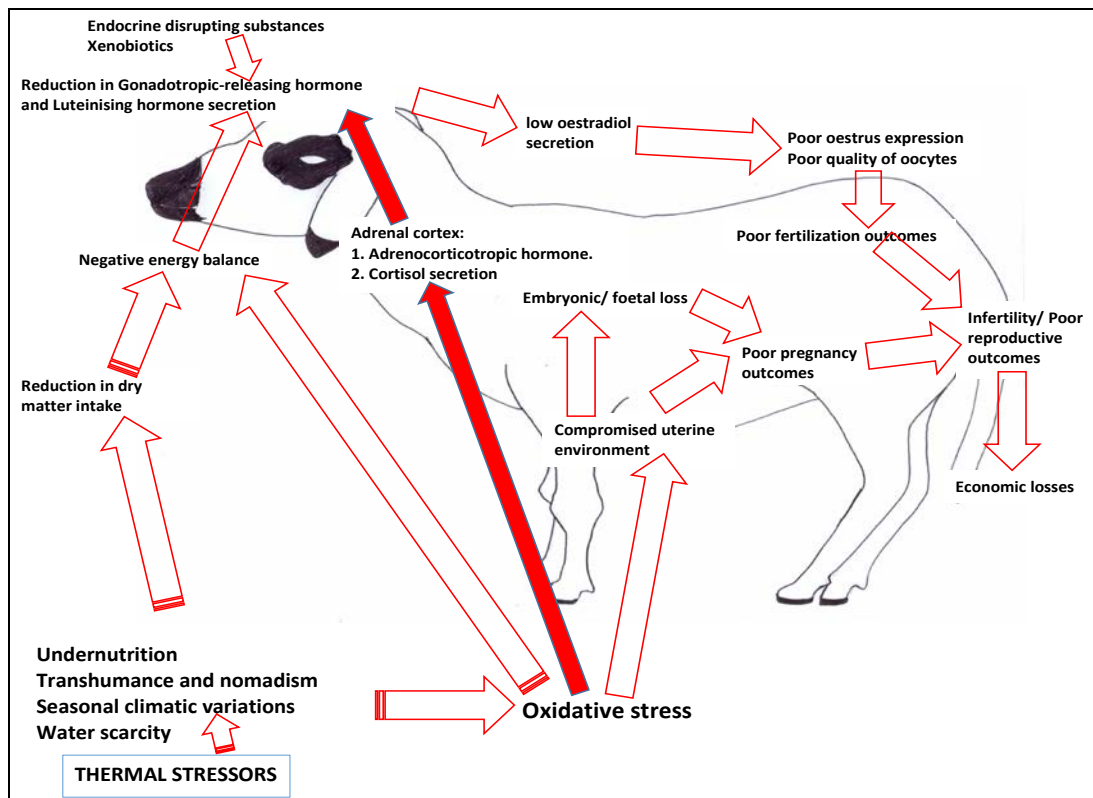


Fig 1: Oxidative stress and its effects on reproductive performance in ewes.

Heat Stress

Heat stress is one of the major factors exerting major negative effects on reproductive performance in ewes and other ruminant species. Although studies on the effects of heat stress on various reproductive parameters have been carried out extensively in cows [44 - 50], its impact on reproductive parameters in the ewes are poorly documented. In a study carried out in Brazil, lower birth and weaning weights in lambs in tropical environments were obtained when compared with those in temperate climate, demonstrating the negative impact of tropical heat stress on reproduction [51]. Heat stress lowers the productive and reproductive efficiency of animals [52]. In other countries, such as Nigeria, sheep productivity is low due to a variety of factors, including the effects of environmental stress, disease burdens, nomadism and undernutrition [53]. Furthermore, it has been shown that elevated rectal temperature, mean maximum and minimum temperatures and relative humidity compounded by feed scarcity during the dry periods of the year are responsible for poor conception rates among ruminants [53]. However, there are no specific information regarding the impact of oxidative stress on conception rates in sheep.

Heat stress reduces libido, fertility, and embryonic survival in ruminants [54]. It also reduces foetal growth and alters the endocrine status of the dam during late gestation [55]. Heat stress changes the endocrine status of sheep to reduce follicular activities and ovulatory mechanism, with a decrease in oocyte and embryo quality [56]. It also results in a decrease in basal metabolism, which subsequently reduce the production of hormones like thyroxine, growth hormone, and glucocorticoids. Heat stress is associated with decrease in electrolyte concentrations such as sodium and potassium in the body and also decrease the production of volatile fatty acids and alters the ratio of acetate/ propionate, and a decline in ruminal pH [55]. Heat stress during summer months has also been reported to reduce the duration and intensity of oestrus in cows, resulting in fewer successful inseminations and poor

insemination outcomes [56, 57]. Heat stress during summer in cows affects the hypothalamic-gonadal axis by reducing the luteinising hormone (LH) levels, including its pulse amplitude and frequency [58, 59]; reduced estradiol secretion by dominant follicle and may demonstrate similar effects in ewes [60, 61, 62]. It lowers plasma inhibin concentration and a relatively higher plasma concentration of follicle stimulating hormone (FSH) levels, resulting in reduced folliculogenesis and poor oestrus behaviour [61]. The reports on the effects of heat stress on plasma progesterone are somewhat inconsistent in cyclic ewes and dairy cows; and were associated with the differences between the rate of luteal production and the rate of hepatic metabolism, which are both affected by changes in dry matter intake [56, 63, 64, 65, 66].

Heat stress in summer lowers the progesterone levels during the luteal phase, which results in poor follicular development, abnormal oocyte maturation and causes embryonic mortality [56, 67]. Although the mechanisms through which heat stress alters the concentrations of circulating reproductive hormones are not known, it has been postulated that increased corticosteroid secretion may be involved through its inhibition of GnRH and, thus, LH secretion [59]. Heat stress is an important factor in gametes and embryonic development and the interaction of such gametes during fertilisation occurs in ruminants. Spermatogenesis is a temperature-sensitive process where temperatures below the normal body temperatures are required. Oocyte development is also influenced by temperature [68]. Heat stress influences the uterine environment through increased uterine temperature, reduced blood flow and reduced endometrial prostaglandin secretions, which result in the inhibition of embryonic development, poor implantation, and premature luteolysis and increase in early embryonic loss, and in overall reduction in the proportion of successful inseminations [56, 69]. Heat stress is reported to affect follicular development through the delay in follicle selection and lengthening of the follicular wave, which in-turn affect the overall oocyte quality and follicular steroidogenesis

[46, 70, 71, 72, 73]. The reduction in food intake occasioned by heat stress and lowered body metabolic activities affects the overall energy balance, resulting in lower LH secretion and decreased dominant follicular size, which may contribute to anovulation in postpartum dairy cows and increase in infertility [56].

A study found the percentage of fertilized ova of 40.7% was significantly lower in ewes subjected to higher temperatures of 32°C than the ewes in the control with fertilized ova of 94.2% [16]. The study also found higher percentage of 55.6% of morphologically-abnormal ova, from 3.8% in the control. The zygote was found to be most sensitive to the harmful effects of high ambient temperature during the initial stages of cleavage before implantation in the uterus [16].

Other studies have associated high ambient temperature with decrease in feed intake efficiency and utilization, disturbances in water, protein, energy and mineral balances, enzymatic reactions, hormonal secretions and blood metabolites, which also impact negatively on reproductive performance [35, 37, 52, 55, 74, 75, 76]. Elevated ambient temperature negatively affects the suprachiasmatic nucleus (SCN) in the brain, which impacts its ability to regulate the circadian and seasonal rhythms of most biological functions, and reproduction, including the phasic and tonic release of hormones, oestrus and gonadal size [37]. Adverse ambient temperature disrupts endocrine function by suppressing oestrus behaviour through a decrease in GnRH/LH pulse frequency and amplitude, with deleterious consequence on fertility in ewes [77]. It was also found in another study that heat-stressed pregnant ewes had lambs with lower birth weight of 3.18 kg., when compared with ranged pregnant ewes (4.57 kg.), while those on restricted feeding had 4.16 kg and dwarfed lambs among the heat-stressed ewes, irrespective of their level of nutrition [78].

Another study showed that the effects of heat stress on the reproductive function in both female and male ruminants are dependent on its severity [79, 80]. Such effects include reduced intensity of behavioural oestrus and low fertility in females, compromised sperm output and increased sperm abnormalities in males. Cows subjected to heat stress had reduced fertility due to changes in endocrine activities, such as reduced estradiol secretion and changes in plasma progesterone concentrations. Such changes reduce the follicular activities and alter the ovulatory mechanisms, leading to a decrease in oocyte and embryo quality, and the entire uterine environment with concomitant effect on implantation [56]. They reported that lactation increases the sensitivity of females to heat stress because of the associated increase in heat production during lactation, occurring due to high metabolic activities. It has been reported that heat stress generally impairs reproductive performance and immune response in livestock [74].

In assisted-reproductive techniques in animals, the production of embryos through superovulation is reduced during heat stress, affecting the success rates of *in vitro* fertilisation (IVF). This occurs as a result of reduced super ovulatory response, lower fertilization rate and reduced embryo quality and viability [78, 79]. The effect of heat stress on the reproductive processes in ewes, essential for embryonic development causes reduced oestrus behaviour expression, changes in follicular development, limiting oocyte performance, and impeding overall embryo growth in the dam [81].

Heat stress has also been associated with compromised passive and cell-mediated immunity in ewes exposed to heat stress during the dry period and fed on their dams' colostrum

[65, 81]. Thus, may also have similar impact on kids, born to sheep exposed to heat stress. Heat stress significantly increase in the packed cell volume (PCV) ($P < 0.05$) but does not significantly alter the total serum thyroxine and triiodothyronine levels in sheep, exposed to heat stress conditions [82]. The rise in PCV is considered as an adaptation mechanism for cooling.

Cold Stress

Cold stress is associated with lower environmental temperatures below the thermo neutral zone when the difference in core body temperature in animals is considerably higher than the environmental temperature. Cold stress has been reported to exert minimal negative effects on reproduction in animals [55]. It is associated with a general reduction in body activities due to lower body temperature. It increases basal metabolism as a measure of generating heat to maintain the body temperature [83]. Increase in metabolic activities in the body results in increased generation of ROS which induce oxidative stress with its consequential adverse effects on reproductive performance. Cold stress may lower conception and pregnancy rates in ewes as reported in heifers [84].

Multiple Environmental Stressors

Multiple environmental stressors, including seasonal effects, also exert deleterious effects on reproduction in ewes. It has been reported that lower reproductive values in ewes subjected to multiple environmental stressors have been obtained, when compared to the control group [43]. There was a significant difference ($p < 0.05$) in the percentages of ewes exposed to multiple environmental stressors (41.7%) and the control group with 66.67%. The oestrus duration (in hours) was significantly higher in the control ewes (32 hours) than ewes in multiple-stressed group (14.4 hours). The conception and lambing rates also dropped significantly from 83.3% in the control group to 50% for the multiple-stressed group. The birth weight was not significantly different between the control and the multiple stressed group. The above study showed poor reproductive performance values in ewes exposed to multiple environmental stressors such as thermal, nutritional and walking stress. Sheep in Nigeria are commonly raised under similar conditions especially in the dry season of year when feed supply is inadequate and animals are walked for long distances in search for food.

Seasonal Effects

A study compared the outcomes of Santa Ines ewes in Brazil under dry and rainy season conditions and found that there was no difference ($P > 0.05$) in expression of oestrus (86.8 and 98.3%), pregnancy rate (86.8 and 90.9%), lambing (76.3 and 83.6%), abortion (9 and 7.4%), fecundity (86.8 and 96.3%), birth (103.4 and 110.8%), abortion rates (9.0% and 7.4%) and prolificacy (113.7 and 115.2%) [85]. However, lamb mortality differs significantly ($P > 0.05$), 26.6% for the dry season group A, and 49% for rainy season group B. Ewes raised in the dry season (group A) had higher average live weight at the end of the reproductive season and during the final third of pregnancy ($P < 0.05$) than those raised in the rainy season (group B). The average daily weight gain was higher for ewes raised in the dry season (98.3 ± 0.009) than in the weight season (8.3 ± 0.008). The live weight during the final third of pregnancy was higher in the dry season group (49.0 ± 1.07) than in the rainy season group (41.4 ± 0.84). The other parameters were not significantly different: post-

partum live weight gain of 39.7 ± 0.98 for dry season group and 36.0 ± 0.78 for the rainy season group, lamb live weight at birth of 3.2 ± 0.12 for dry season group and 3.0 ± 0.09 for the rainy season group, male lamb live weight from singletons (kg) of 3.3 ± 0.13 for dry season group and 3.1 ± 1.0 for the rainy season group, females from singletons (kg) 3.5 ± 0.13 for dry season group and 3.0 ± 0.09 for the rainy season group, males lamb weight from twins live weight (kg) 2.9 ± 0.08 for dry season group and 2.4 ± 1.0 for the rainy season group, and females lamb weight from twins live weight (kg) of 2.2 ± 0.092 for dry season group and 2.6 ± 0.091 for the rainy season group.

The above studies highlight the variations in the negative effects of oxidative stress associated with environmental stressors, which are common problems facing sheep production in developing tropical countries such as Nigeria. The practice of extensive management system through walking of animals for long distances, poor nutrition especially during the late dry season, high thermal stress and solar radiation are important stressors compromising reproductive performance of sheep. Studies in Nigeria regarding the impact of oxidative stress on reproductive performance in ewes have not been adequately documented.

Concluding Remarks

Oxidative stress remains an important condition undermining reproductive performance in sheep. It is a silent cause of economic loss through poor oestrus behaviour and mating, poor fertilisation and overall pregnancy outcomes. It exerts and retards lambs' growth and development, including compromise of the ability of sheep to resist diseases and other stressful conditions. There are measures, such as the use of antioxidants, aimed at minimizing the effects of oxidative stress, posed by environmental stressors on reproduction. Such measures may be beneficial in ameliorating the negative impact of oxidative stress and may enhance sheep productivity.

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Conflict of Interest

The authors declare that there is no conflict of interest associated with this review article.

References

1. Salami A, Kamara AB, Brixiova Z. Smallholder agriculture in East Africa: trends, constraints and opportunities. African Development Bank. Working Paper No 2010, 105.
2. Ibrahim H. International Livestock Research Institute (ILRI) Small Ruminant Production Techniques. ILRI Manual 1998, 3. Nairobi, Kenya.
3. Yoo HS. Infectious causes of reproductive disorders in cattle. *Journal of Reproduction and Development* 2010;56(Suppl):S53-S60.
4. Bonneville-Hébert A, Bouchard E, Tremblay, DD, Lefebvre R Effect of reproductive disorders and parity on repeat breeder status and culling of dairy cows in Quebec. *Canadian Journal of Veterinary Research* 2011;75(2):147-151.

5. Khair A, Alam MM, Rahman AKMA, Islam MT, Azim A, Chowdhury EH. Incidence of reproductive and production diseases of cross-bred dairy cattle in Bangladesh. *Bangladesh Journal of Veterinary Medicine* 2013;11(1):31-36.
6. Banerjee P, Bhattacharya J. Impact of oxidative stress on infertility, with emphasis on infertility management strategies. *Global Journal of Fertility and Research* 2019;4(1):10-18.
7. Dawane JS, Pandit VA. Understanding redox homeostasis and its role in cancer. *Journal of Clinical and Diagnostic Research* 2012;6(10):1796-1802.
8. Sorelle DN, Ferdinard N, Narcisse VB, Tchoumboue. Medicinal plants and female reproductive disorders due to oxidative stress. *Archive of Veterinary Science and Medicine* 2019;2(4):58-73. Doi: 10.26502/avsm.009.
9. Sies H, Berndt C, Jones DP. Oxidative stress. *Annual Review of Biochemistry* 2017;86:715-748. www.annualreview.org.
10. Banerjee BD, Seth V, Ahmed RS. Pesticide-induced oxidative stress: perspective and trends. *Reviews on Environmental Health* 2010;16(1):1-40. Doi: <https://doi.org/101515/REVETH.2001.16.1>.
11. Celi P. The role of oxidative stress in small ruminants' health and production. *Revista Brasileira de Zootecnia* 2010;39:348-363, (supl. especial).
12. Agarwal A, Rana M, Qiu E, AlBunni H, Bui AD, Henkel R. Role of oxidative stress, infection and inflammation in male infertility. *Andrologia* 2018;50:e13126. <https://doi.org/10.1111/and.13126>.
13. Chauhan SS, Celi P, Ponnampalam EN, Leury BJ, Liu F, Dunshea FR. Antioxidant dynamics in the live animal and implications for ruminant health and product (meat/milk) quality: role of vitamin E and selenium. *Animal Production Science* 2014a;54:1525-1536.
14. Zhong R, Zhou D. Oxidative stress and the role of natural plant-derived antioxidants in animal reproduction. *Journal of Integrative Agriculture* 2013;12(10):1826-1838.
15. Agarwal A, Saleh RA, Bedaiwy MA. Role of reactive oxygen species in the pathophysiology of human reproduction. *Fertility and Sterility* 2003;79(4):829-843.
16. Dutta S, Majzoub A, Agarwal A. Oxidative stress and sperm function: a systematic review on evaluation and management. *Arab Journal of Urology* 2019;17(2):87-97.
17. Nowicka-Bauer K, Brett N. Molecular changes induced by oxidative stress that impair human sperm motility. *Antioxidants* 2020;9:134. Doi:10.3390/antiox9020134. www.mdpi.com/journal/antioxidants.
18. Agarwal A, Gupta S, Sharma RK. Role of oxidative stress in female reproduction. *Reproductive Biology and Endocrinology* 2005;3:28.
19. Parijatham S, Saikumar P, Chandra SE. Importance of oxidative stress marker in healthy females undergoing assisted reproductive technology. *Journal of Environmental and Analytical Toxicology* 2015;5:331. Doi: 10.4172/2161-0525.1000331.
20. Mutinati M, Piccinno M, Roncetti M, Campanile D, Rizzo A, Sciorsci R. Oxidative stress during pregnancy in the sheep. *Reproduction of Domestic Animals* 2013;48(3):353-357.
21. Agarwal A, Said TM, Bedaiwy MA, Banerjee J, Alvarez JG. Oxidative stress in an assisted reproductive technique setting. *Fertility and Sterility* 2006b;86(3):503-512.

22. Armanios M, Alder JK, Parry EM, Karim B, Strong MA, Greider CW. Short telomeres are sufficient to cause the degenerative defects associated with aging. *The American Journal of Human Genetics* 2009;85:823-832.
23. Desai N, Sabanegh E, Kim T, Agarwal A. Free radical theory of ageing: Implications in male infertility. *Urology* 2010;75(1):14-19.
24. Showell MG, Brown J, Yazdani A, Stankiewicz MT, Hart RJ. Re: Antioxidants for male subfertility. *European Urology* 2011;62:562-567.
25. Rathwa SD, Vasava AA, Pathan MM, Madhira SP, Patel YG, Pande AM. Effect of season on physiological, biochemical, hormonal, and oxidative stress parameters of indigenous sheep. *Veterinary World* 2017;10(6):650-654. www.veterinaryworld.org/Vol.10/June-2017/13.pdf.
26. Aitken RJ. Impact of oxidative stress on male and female germ cells: implications for fertility. *Reproduction* 2020;159:R159-R201.
27. Pedernera M, Celi P, García SC, Salvin HE, Barchia I, Fulkerson WJ. Effect of diet, energy balance and milk production on oxidative stress in early-lactating dairy cows grazing pasture. *The Veterinary Journal* 2010;186:352-357.
28. Salifu S. Reproductive performance of Djallonké sheep in the northern region of Ghana. A Thesis submitted to the School of Graduate Studies, Kwame Nkrumah University of Science and Technology, Kumasi, in fulfilment of the requirements for the Award of Master of Philosophy (Reproductive Physiology) Degree, College of Agriculture and Natural Resources, Department of Animal Science, Faculty of Agriculture, Kwame Nkrumah University of Science and Technology, Kumasi, Ghana 2014.
29. Ribas-Maynou Y, Salas H. The relationship between sperm oxidative stress alterations and IVF/ICSU outcomes: a systematic review from nonhuman mammals. *Biology* 2020;9:178. Doi: 10.3390/biology9070178.
30. Menezo Y. Antioxidants to reduce sperm DNA fragmentation: an unexpected adverse effect. *RBM Online* 2007;14(4):418-421.
31. Fang Y, Zhong R. Effects of oxidative stress on spermatozoa and male infertility. *In: Free Radical Medicine and Biology (working title)* 2019. Doi: <http://dx.doi.org/105772/intechopen.8685>.
32. Shoorei H, Khaki A, Khaki AA, Hemmati AA, Moghimian M, Shokohi M. The ameliorative effect of carvedilol on oxidative stress and germ cell apoptosis in testicular tissue of adult diabetic rats. *Biomedicine and Pharmacotherapy* 2019;111:568-578. <https://doi.org/10.1016/j.biopha.2018.12.054>.
33. Shahat AM, Rizzoto G, Kastelic JP. Amelioration of heat stress-induced damage to testes and sperm quality. *Theriogenology* 2020;158:84-96. <https://doi.org/10.1016/j.theriogenology.2020.08.034>.
34. Chauhan SS, Celi P, Leury BJ, Clarke IJ, Dunshea FR. Dietary antioxidants at supranutritional doses improve oxidative status and reduce the negative effects of heat stress in sheep. *Journal of Animal Science* 2014b;92:3364-3374.
35. Marai IFM, El-Darawany AA, Fadiel A, Abdel-Hafez MAM. Physiological traits as affected by heat stress in sheep: A review. *Small Ruminant Research* 2006;71(1):1-12.
36. Bansal AK, Bilaspuri GS. Impacts of oxidative stress and antioxidants on semen functions. *Veterinary Medicine International* 2011;2011:1-7.
37. Visioli F, Hagen TM. Antioxidants to enhance fertility: Role of eNOS and potential benefits. *Pharmacological Research* 2011;64:431-437.
38. Rodriguez-Gonzalez GL, Reyes-Castro LA, Vega CC, Boeck L, Ibanez C, Nathanielsz PW, *et al.* Accelerated aging of reproductive capacity in male rat offspring of protein-restricted mothers is associated with increased testicular and sperm oxidative stress. *Age* 2014;36:9721. Doi: 10.1007/s11357-014-9721-5.
39. Al-Musawi JE, Hassan SA, Muhammad SF. Effect of cold stress on some blood parameters of sheep and goats. *International Journal Science and Research* 2015, 2319-7064. paper ID: ART20164378. Doi: <https://doi.org/10.2127/ART2016437>.
40. Krishnan G, Bagath M, Pragna P, Vidya MK, Aleena J, Archana PR, *et al.* Mitigation of the heat stress impact in livestock reproduction. *In: Theriogenology*, Carreira, R. P. Editor, Intechopen. Available from: <https://www.intechopen.com/books/theriogenology> 2017. <http://dx.doi.org/10.5772/intechopen.69091>. Accessed 30 11 2019, 5.30 p.m.
41. Olanrewaju RM, Tilakasiri SL, Adeleke EA. Effects of climate on chicken production in Ilorin, Kwara State, Nigeria. *The Journal of Agricultural Sciences* 2016;11(2):88-96.
42. Silanikove N. Effects of heat stress on the welfare of extensively managed domestic ruminants. *Livestock Production Science* 2000;67:1-18.
43. Sejian V, Maurya VP, Naqvi SMK. Effect of multiple stresses (thermal, nutritional, and walking stress) on the reproductive performance of Malpura ewes. *Veterinary Medicine International* 2012. Article ID 471760, 5 pages.
44. Zeron Y, Ocheretny A, Kedar O, Borochoy A, Sklan D, Arav A. Seasonal changes in bovine fertility: relation to developmental competence of oocytes, membrane properties and fatty acid composition of follicles. *Reproduction* 2001;121:447-454.
45. Wolfenson D, Sonego H, Bloch A, Shaham-Albalancy A, Kaim M, Folman Y, *et al.* Seasonal differences in progesterone production by luteinized bovine thecal and granulosa cells. *Domestic Animal Endocrinology* 2002;22(2):81-90.
46. Sönmez M, Demirci E, Türk G, Gür S. Effect of season on some fertility parameters of dairy and beef cows in Elaz Province. *Turkish Journal of Veterinary and Animal Science* 2005;29:821-828.
47. Villa-Mancera A, Méndez-Mendoza M, Huerta-Crispín R, Vázquez-Flores F, Córdova-Izquierdo A. Effect of climate factors on conception rate of lactating dairy cows in Mexico. *Tropical Animal Health and Production* 2011;43(3):597-601.
48. Dunshea FR, Leury BJ, Fahri F, DiGiacomo K, Hung A, Chauhan S, *et al.* Amelioration of thermal stress impacts in dairy cows. *Animal Production Science* 2013;53(9):965-975.
49. El-Wishy AB. Fertility of Holstein cattle in a subtropical climate of Egypt. *Iranian Journal of Applied Animal Science* 2013;3(1):45-51.
50. Schüller LK. Influence of heat stress on the reproductive performance of dairy cows in the moderate climate of the temperate latitude. *Aus der Tierklinik für Fortpflanzung des Fachbereichs Veterinärmedizin der Freien Universität*

- Berlin. Inaugural Dissertation zur Erlangung des Grades eines Doktors der Veterinärmedizin an der Freien Universität Berlin. Retrieved from: http://www.diss.fu-berlin.de/diss/servlets/MCRFileNodeServlet/FUDISS_derivate_000000016979/Schueller_online.pdf 2014, accessed 1/9/2016, 8.05 a.m.
51. Combellas J. Production and reproduction parameters of tropical sheep breeds in improved production systems. *Tropical Animal Production* 1980;5(3):266-272.
 52. Fuquay JW. Heat stress as it affects animal production. *Journal of Animal Science* 1981;52(1):164-174.
 53. Zakari AY, Molokwu ECI, Osori DIK. Effects of rectal and ambient temperatures and humidity on conception rates. *Theriogenology* 1981;16(3):331-336.
 54. Singh SP, Kumar A, Sourya N. Effects of heat stress on animal reproduction. *International Journal of Fauna and Biological Sciences* 2021;8(2):16-20.
 55. Collier RJ, Beede DK, Thatcher WW, Israel LA, Wilcox CJ. Influences of environment and its modification on dairy animal health and production. *Journal of Dairy Science* 1982;65(11):2213-2227.
 56. de Rensis F, Scaramuzzi RJ. Heat stress and seasonal effects on reproduction in the dairy cow – a review. *Theriogenology* 2003;60(6):1139-1151.
 57. Younas M, Fuquay JW, Smith AE, Moore AB. Estrus and endocrine responses of lactating Holsteins to forced ventilation during summer. *Journal of Dairy Science* 1993;76:430-434.
 58. Wise ME, Armstrong DV, Huber JT, Hunter R, Wiersma F. Hormonal alterations in the lactating dairy cow in response to thermal stress. *Journal of Dairy Science* 1988;71:2480-2485.
 59. Gilad E, Meidan R, Berman A, Graber Y, Wolfenson D. Effect of heat stress on tonic and GnRH-induced gonadotrophin secretion in relation to concentration of oestradiol in plasma of cyclic cows. *Journal of Reproduction and Fertility* 1993;99:315-321.
 60. Wolfenson D, Lew BJ, Thatcher WW, Graber Y, Meidan R. Seasonal and acute heat stress effects on steroid production by dominant follicles in cows. *Animal Reproduction Science* 1997;47(1-2):9-19.
 61. Wolfenson D, Thatcher WW, Badinga L, Savio JD, Meidan R, Lew BJ, *et al.* The effect of heat stress on follicular development during the estrous cycle dairy cattle. *Biology of Reproduction* 1995;52:1106-1113.
 62. Wilson SJ, Marion RS, Spain JN, Spiers DE, Keisler DH, Lucy MC. Effect of controlled heat stress on ovarian function in dairy cattle: I. Lactating cows. *Journal of Dairy Science* 1998;81(8):2124-2131.
 63. Sheikheldin MA, Howland BE, Palmer WM. Effects of heat stress on serum progesterone in cyclic ewes and on progesterone and cortisol response to ACTH in ovariectomised ewes. *Journal of Reproduction and Fertility* 1988;84:521-529.
 64. Bell AW, McBride BW, Slepatis R, Early RJ, Currie WB. Chronic heat stress and prenatal development in sheep: I. conceptus growth and maternal plasma hormones and metabolites. *Journal of Animal Science* 1989;67:3289-3299.
 65. Sevi A, Caroprese M. Impact of heat stress on milk production, immunity and udder health in sheep: a critical review. *Small Ruminants Research*. 2012;107:1-7.
 66. Macias-Cruz U, Gastelum MA, Alvarez FD, Correa A, Diaz R, Meza-Harrera CA, *et al.* Effects of summer heat stress on physiological variables, ovulation and progesterone secretion in Pelibuey ewes under natural outdoor conditions in an arid region. *Animal Science Journal* 2016;87:354-360. Doi: <https://doi.org/10.1111/asj.12430>.
 67. Thwaites CJ. Embryo mortality in the heat stressed ewe. I: the influence of breed. *Journal of Reproduction* 1967;14:5-14.
 68. Rivera RM, Al-Katanani YM, Paula-Lopes FF, Hansen PJ. Short Communication: Seasonal effects on development of bovine embryos produced by *in vitro* fertilization in a hot environment. *Journal of Dairy Science* 2000;83:305-307.
 69. Rivera RM, Hansen PJ. Development of cultured bovine embryos after exposure to high temperature in the physiological range. *Reproduction* 2001;121:107-115.
 70. Torres-Júnior JRS, Pires MFA, de Sa WF, Ferreira AM, Viana JHM, Camargo LSA, *et al.* Effect of maternal heat-stress on follicular growth and oocyte competence in *Bos indicus* cattle. *Theriogenology* 2008;69:155-166.
 71. Teke B, Akdag F. The effect of heat stress on some reproductive traits in Jersey cows under semi-humid conditions in Turkey. *Bulgarian Journal of Agricultural Science* 2012;18(4):506-510.
 72. Paula-Lopes FF, Lima RS, Risolia PHB, Ispada J, Assumpção MEOA, Visintin JA. Heat stress induced alteration in bovine oocytes: functional and cellular aspects. *Animal Reproduction* 2012;9(3):395-403.
 73. Verma KK, Prasad S, Mohanty TK, Kumaresan A, Layek SS, Patbandha TK *et al.* Effect of short-term cooling on core body temperature, plasma cortisol and conception rate in Murrah buffalo heifers during hot-humid season. *Journal of Applied Animal Research* 2016;44(1):281-286.
 74. Nardone A, Ronchi B, Lacetera N, Ranieri MS, Bernabucci U. Effects of climate changes on animal production and sustainability of livestock systems. *Livestock Science* 2010;130(1):57-69.
 75. Sejian V, Maurya VP, Naqvi SMK. Adaptability and growth of Malpura ewes subjected to thermal and nutritional stress. *Tropical Animal Production and Health* 2010a;42(8):1763-1770.
 76. Sejian V, Maurya VP, Naqvi SMK. Adaptive capability as indicated by endocrine and biochemical responses of Malpura ewes subjected to combined stresses (thermal and nutritional) in a semi-tropical environment. *International Journal of Biometeorology* 2010b;54(6):653-661.
 77. Dobson H, Fergani C, Routly JE, Smith RF. Effects of stress on reproduction in ewes. *Animal Reproduction Science* 2012;130(3):135-140.
 78. Brown DE, Harrison PC, Hinds FC, Lewis JA, Wallace MH. Heat Stress Effects on foetal development during late gestation in the ewe. *Journal of Animal Science* 1977;44:442-446.
 79. Hansen PJ, Drost M, Rivera RM, Paula-Lopes FF, Al-Katanani YM, Krininger CE, *et al.* Adverse impact of heat stress on embryo production: causes and strategies for mitigation. *Theriogenology* 2001;55(1):1-103.
 80. Hansen PJ, Fuquay JW. Stress in Dairy Animals. Heat Stress: Effects on Reproduction. Reference Module in Food Science Encyclopedia of Dairy Sciences (Second Edition), Research Gate 2015, 567-574.
 81. Van Wettere HEJ, Kind KL, Gatford KL, Swinbourne AM, Leu ST, Hayman PT, *et al.* Review of the impact of heat stress on reproductive performance of sheep. *Journal*

- of Animal science and Biotechnology 2021;12:26. Doi: <https://doi.org/10.1186/s40104-020-00537-z>.
82. Al-Haidary AA. Physiological responses of Niamey sheep to heat stress challenge under semi-arid environments. *International Journal of Agriculture and Biology* 2004;6(2):307-309.
 83. Cannon B, Nedergaard J. Nonshivering thermogenesis and its adequate measurement in metabolic studies. *The Journal of Experimental Biology* 2011;214:242-253.
 84. Chebel RC, Braga FA, Dalton JC. Factors affecting reproductive performance in holstein heifers. *Articles in Animal Reproduction Science* 2007;101(3-4):208-224.
 85. Moura ACB, Teixeira PPM, Coutinho LN, Paz CC, Santos VJC, Soares FN, *et al.* Reproductive performance of Santa Inês ewes during dry and rainy seasons in eastern Amazon. *Animal Reproduction* 2014;11(1):44-48.